

Review

Genomic Insights into Cardiomyopathies: A Comparative Cross-Species Review

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Abstract: In the global human population, the leading cause of non-communicable death is cardiovascular disease. It is predicted that by 2030, deaths attributable to cardiovascular disease will have risen to over 20 million per year. This review compares the cardiomyopathies in both human and non-human animals and identifies the genetic associations for each disorder in each species/taxonomic group. Despite differences between species, advances in human medicine can be gained by utilising animal models of cardiac disease; likewise, gains can be made in animal medicine from human genomic insights. Advances could include undertaking regular clinical checks in individuals susceptible to cardiomyopathy, genetic testing prior to breeding, and careful administration of breeding programmes (in non-human animals), further development of treatment regimes, and drugs and diagnostic techniques.

Keywords: cardiomyopathy; human; canine; cross-species; cardiovascular disease; genetics

1. Introduction

The cardiomyopathies (CMs) are a diverse group of cardiac disorders of the myocardium first described in the 1950s, usually exhibiting clinically unexplained hypertrophy or dilation of the ventricular walls and/or septum which can be either solely confined to the heart, or one part of a more generalised systemic disorder [1–4]. Cardiomyopathies are present in all populations and age groups and can lead to heart failure and death. Types of cardiomyopathy include: hypertrophic cardiomyopathy (HCM); dilated cardiomyopathy (DCM); restrictive cardiomyopathy (RCM); arrhythmogenic right ventricular cardiomyopathy (ARVC); and unclassified cardiomyopathies [5]. There is a range of mammalian species that can also develop the same cardiomyopathies either naturally or via induced disease processes. In addition, there are cardiomyopathies that are unique to a species/taxonomic group, such as great ape cardiomyopathy [6]. Although no genetic links have been published to date in relation to the great ape cardiomyopathy, collaborative work is presently underway throughout Europe under “The Ape Heart Project” [7]. Both natural and induced disease can act as models for other species, including humans, but naturally occurring cardiomyopathy (CM) in agricultural, working, and companion animals is also of clinical and financial concern in its own right.

The aetiology of the cardiomyopathies is complex, as causation can be both acquired and genetic in origin and often unknown. HCM is usually genetically inherited, with a prevalence of about 1:500 [5]. DCM is genetic in about one-third of cases [8] and can also be caused by external factors such as alcohol, toxins, drug abuse, viral infections, and pregnancy. Many (25%) of the mutations associated with this form are in the large titin gene [9]. RCM is rare genetically and can be caused as a result of connective

tissue disorders, sarcoidosis, amyloidosis and haemochromatosis, eosinophilic heart disease, or as a result of radiation treatment [10], but has also been linked to several genetic mutations [11,12]. ARVC has been linked to several genes, but mutations in genes affecting the desmosomes occur most frequently within this disorder. Linkage of CM to a genetic locus was first reported in 1989 [13], and in 1990 the first causative gene mutation (in cardiac beta (β)-myosin heavy chain; *MYH7*/ β *MHC*) was reported [14]. The genetics of the CMs is complex due to both phenotype and genotype heterogeneity over a wide range of severities. Symptoms can vary from none to very extreme, and risk of death may vary from almost zero with a normal lifespan to early infant death and clinical progression, and from no measurable progression to rapid and early symptoms. To complicate matters further, the transmission is often autosomally dominant but with variable penetrance, and there are affected genes on the X chromosome (recessive inheritance) and also within the mitochondrial genome which are maternally inherited. This review gives an oversight into what has been published for human and non-human animals to date. The search terms throughout included cardiomyopathy alongside a species name using both the Latin and common names.

2. Hypertrophic Cardiomyopathy (HCM)

In the 2014 European Society of Cardiology (ESC) guidelines on diagnosis and management of hypertrophic cardiomyopathy [15], the disorder is defined by “the presence of increased left ventricular wall thickness that is not solely explained by abnormal loading conditions”. It is characterised by left ventricular hypertrophy (without dilation) and abnormally large and misaligned myocytes localised to the interventricular septum; many patients develop dynamic obstruction to left ventricular outflow and increased fibrosis which leads to diastolic heart failure [5,16]. HCM is the most common form of CM, which has a reported prevalence of 0.02%–0.23% in adults [15,16]. There have been HCM-associated mutations in over 20 genes associated with the sarcomere, myofilament, Z-discs, and calcium handling in humans [17].

Most of the HCM mutations (up to 80% [18]) come from the beta myosin heavy chain (*MYH7* 25%–40%) and cardiac myosin binding protein C genes (*MYBPC3*, 25%–40%), which are genes of the sarcomere/myofilament proteins that generate the force for myocyte contraction. The cardiac troponin T genes *TNNT2* and *TNNi3* contribute a further 10% more cases and it is well established that genes of the Z-disc and calcium-handling genes, amongst many others, contribute less commonly to the causes of CM [16,18].

In addition to its occurrence in people, HCM occurs naturally in both cats [19] and dogs [20,21]. It is more common in cats than dogs, accounting for <1% of canine cardiovascular diagnoses [20], but it has been diagnosed in up to 14.6% of apparently healthy cats [22]. Feline hypertrophic cardiomyopathy appears to have a breed predisposition, as does canine DCM and is, reportedly, the most common cardiac disorder in cats and has shown remarkable similarities to human HCM [19]. The familial predisposition is most obvious in Maine Coon, Ragdoll, and British and American Shorthair breeds [23–25]. Separate genetic mutations have been identified as causal of HCM in the Maine Coon and Ragdoll breeds within the *MYBPC3* gene [24,26] but there are also affected individuals within the Maine Coon breed lacking the mutation, indicating that there are additional causes of the disease to be established [27]. *MYBPC3* is a gene with mutations associated with human HCM [28]; thus, in these cases, feline and human HCM may act as relevant models for each other. This is important, considering that an estimated 35% of human cases are caused by *MYBPC3* mutations [16]. Higher concentrations of cTnI have also been observed in some breeds of cats with HCM [25,29], but no mutations have been observed in that gene in Maine Coons or British Shorthairs [25], whereas mutations in this gene have been observed in humans. Pigs also appear to have a heritable form of HCM, but no specific genetic associations have been discovered to date [30].

There are fewer available induced models of HCM compared to DCM. The primary model type being transgenic models, generally based on genes associated with human HCM in order to create the HCM phenotype. Transgenic and naturally occurring strains of mice [31], rabbits [32],

and hamsters [33] have been used as models of HCM. Some strains of hamsters—including BIO14.6 and TO-2—develop HCM and DCM showing mutations in the delta-sarcoglycan gene, however, the phenotype differs a little from humans in that it shows augmented necrosis [34,35].

3. Dilated Cardiomyopathy (DCM)

Dilated cardiomyopathy is defined by left ventricular chamber enlargement and systolic dysfunction. Enlargement of the cavity and thinning and weakening of the walls of the left ventricle generally progresses to the right ventricle over time. The inability to pump properly leads to heart failure and clot formation in the heart in some species [36]. DCM is estimated to have a prevalence of 0.036% in humans but this is thought to be an underestimate and, in common with the other cardiomyopathies, the incidence is increasing with better testing and recognition [37–39]. To date, there have been mutations in over 50 genes associated with human DCM [40].

Naturally occurring spontaneous DCM exists in several species including dogs [41], cats [42], cattle [43–45], turkeys [46,47], and chickens [48]. Feline DCM is frequently associated with a lack of dietary taurine, and since this discovery commercial pet foods are now supplemented with taurine. Feline DCM is now rare [49,50]. Despite this, there are instances of feline DCM that are not related to taurine deficiency [42,50]. These instances are currently unexplained, but it is possible that, as with other species, the underlying cause of these cases is genetic. No genetic loci have been associated with feline DCM, but evidence for a genetic involvement in the development of feline DCM has been demonstrated [51].

Naturally occurring canine DCM is common—10% of dogs that died from cardiac disease within the insured Swedish dog population had a diagnosis of DCM [20]. Whilst canine DCM is common, there are some breeds that are particularly susceptible to developing the disease [20]. There are thought to be several sub-types of canine DCM which may reflect those seen in human DCM [52]. An example of this is juvenile DCM that occurs in Portuguese water dogs, which may reflect childhood DCM in humans [53,54]. Despite similarities with human DCM, and many studies testing for genetic associations with canine DCM, there have only been 10 loci associated with adult-onset canine DCM [55–59], and recently reviewed by Simpson et al. [52]. There has been an additional locus identified as associated with juvenile-onset canine DCM [60].

DCM in cattle is reported to particularly affect Red Holstein and Simmentaler X Red Holstein individuals [43,44]. It is possible that individuals from other breeds of cattle develop DCM, but within the commercial dairy and beef industries any such disease is likely to be rapidly selected against as it would have an impact on production. DCM in cattle was demonstrated to be inherited in an autosomal recessive pattern [61]. A nonsense mutation in the outer mitochondrial membrane lipid metabolism regulator (*OPA3*) gene has subsequently been demonstrated to be responsible for these incidences of bovine DCM [44].

DCM in turkeys has been associated with unusually low molecular weight cardiac troponin T due to the splicing out of exon 8 [46]. Exon 8 is not normally spliced out of avian and mammalian cardiac troponin T, thus it is considered to be abnormal with probable functional implications contributing to the development of turkey DCM [46]. Exon 8 is also spliced out of cardiac troponin T in wild turkeys [46] and thus likely contributed to the death of the wild turkey [47]. It is of interest that cardiac troponin T is also implicated in humans [62,63] and a mouse model [31]. Chicken DCM has been reported as a naturally occurring disease [48], but also of interest are the *in ovo* chicken models showing that knockdown of the myosins—specifically, embryonic myosin heavy chain (*eMHC*)—results in DCM [64], as does alpha myosin heavy chain (α MHC/*MYH6*) [65] which also affects humans [66,67].

DCM-induced animal models are frequently used in order to understand not only the genetic factors but also the structural and functional aspects of the disease. Several inbred strains of hamster develop DCM [35] and a number of methods are used in different species to induce disease using chemical and mechanical induction, including pacing [68–71].

4. Restrictive Cardiomyopathy (RCM)

Restrictive cardiomyopathy results in dilated atria and stiffening of the ventricles, which leads to restrictive filling and reduced diastolic volume, inefficient pumping, and heart and valve failure. Hypertrophy is typically absent and systolic function is usually unaffected [71]. RCM in children is rare: it accounts for only 2%–5% of childhood cardiomyopathies [72]. Although the incidence of RC is rare, it has a poor prognosis, with 30% of affected patients dying within 5 years of diagnosis [5]. Although RCM is much rarer than DCM, HCM, and ARVC, mutations in several genes have been associated with RCM, including beta myosin heavy chain (*MYH7*) and troponin I [12,73,74]. The RCM-causing mutation was identified in 19 patients (60%). Mutated genes have also been implicated in a number of idiopathic RCM cases. In a recent large cohort, the number of affected individuals showing a genetic change numbered between one and four patients for each gene investigated, *MYH7* (four patients), desmin and filamin C (*DES* and *FLNC*; three patients each), *MYBPC3* and lamin A (*LMNA*; two patients each), titin-cap, troponin I3 cardiac type, troponin T2 cardiac type, tropomyosin 1 (alpha), and lysosomal associated membrane protein 2 (*TCAP*, *TNNI3*, *TNNT2*, *TPM1*, and *LAMP2*; one patient each) [75,76].

RCM has recently been shown to naturally affect cats [77,78]. There have not been any loci associated with feline RCM and from the initial report it may be difficult to identify genetic loci as there does not appear to be familial inheritance because, in this study, individuals were from a range of breeds and were not known to have been related [77]. A mouse model expressing a missense mutation (R193H) in troponin I3 cardiac type (*CTnl*) has been developed [79]. Mice affected by sickle cell anaemia have recently shown symptoms of RCM [80] and this link has also been confirmed in humans [81]. There is some discussion as to whether this is a unique form of cardiomyopathy or RCM, as it is characterised by diastolic dysfunction, left atrial dilation, and normal systolic function [81].

5. Arrhythmogenic Right Ventricular Cardiomyopathy (ARVC)

Arrhythmogenic right ventricular cardiomyopathy (ARVC also called ARVD) is characterised by fibro fatty replacement of the right ventricular myocardium [82]. As the proteins forming the heart scaffolding become defective, muscle cell death occurs and muscle tissue is replaced with fatty and fibrous tissue. The heart walls become thinner and pumping efficiency and cardiac rhythm are affected, leading to heart failure [82]. In humans, ARVC has a reported prevalence of 0.02%–0.10% in the general population and is associated with other disorders such as amyloidosis [18,83]. There have been mutations in several genes identified as associated with human ARVC, in particular, mutations in the five desmosomal genes—desmoplakin, plakophilin 2, desmoglein 2, desmocollin 2, and junction plakoglobin (*DSP*, *PKP2*, *DSG2*, *DSC2*, and *JUP*) [84]. It is thought that around 50% of symptomatic humans have a mutation in one of these five cardiac desmosome genes. Other non-desmosomal genes have also been implicated including transforming growth factor beta 3, transmembrane protein 43, desmin, lamin A, titin, phospholamban, and catenin alpha 3 (*TGFB3*, *TMEM43*, *DES*, *LMNA*, *TTN*, *PLN*, and *CTNNA3*) [85]. In addition, ryanodine receptor 2 (*RYR2*) has been implicated in both human ARVC [86] and in a chronic anthracycline-induced cardiomyopathy in mice [87]. Numerous mutations have been described in relation to causing ARVC including the arrhythmogenic right ventricular dysplasia genes *ARVD3* (14q12–q22) [88], *ARVD4* (2q32.1–q32.3) [89], *ARVD6* (10p14–p12) [90,91], and *ARVD7/ARVC7* (10q22.3) [92].

ARVC naturally affects dogs but has only been widely reported in the Boxer breed [93] and has been suggested as a model of human ARVC. There has been a mutation in the striatin (*STRN*) gene associated with ARVC in the Boxer dog [94]; it is of interest that the same gene has also been associated with DCM in the Boxer [57]. Striatin has not yet been implicated in human ARVC, but could be an area of interest to investigate. There is a syndrome in cattle that has a cardiac element to it that resembles that observed in humans where the cardiac element is ARVC. A mutation in the nuclear factor kappa B subunit 1 (*NFKB1*) gene is associated with this syndrome [95]. It is of interest that a functional polymorphism of *NFKB1* has also been linked to human DCM [96]. Despite identifying ARVC in cats

over 15 years ago [97], and more recently in horses [98,99], there are no reports of genetic associations with feline or equine ARVC in the literature to our knowledge.

6. Mitochondrial, X Linked, and Peripartum Cardiomyopathies

It has been suggested that ~5% of DCMs are X-linked [100]. The X-linked cardiomyopathies are often associated with systemic general disorders such as Fabry's disease, Barth syndrome, and Duchenne and Becker muscular dystrophy, and are most commonly associated with DCM and HCM [5,101–104]. Most of the work is presently in humans and very frequently on families, but the main proteins mutated are tafazzin (G4.5), emerin, lysosome-associated membrane protein 2, XK membrane transport protein, and dystrophin [104–108].

It is likely that mitochondrial genes are often associated with HCM and DCM because the heart is a high user of cellular energy provided by mitochondria. tRNA genes are associated with myoclonic epilepsy with ragged red fibres (MELAS) and mitochondrial encephalopathy with lactic acidosis and stroke-like episodes (MERRF) syndromes, and inheritance is maternally inherited [109]. This category often includes cardiomyopathy caused as a result of general systemic disorders where symptoms are not wholly associated with the heart [5]. HLA-DRB1*0901 allele has also been associated with a number of DCM patients, which suggested that mitochondrial ADP/ATP plays a large role in appropriate functionality of the heart [110]. It has also been shown that mitochondrial mutations are associated with changes in cardiomyopathy forms. The Mt8348A→G mutation in the mitochondrial tRNA(Lys) gene has shown a phenotypic alteration from hypertrophic to severe dilated cardiomyopathy [111], but so far only one case has been shown and therefore more patients need to be analysed. The role of mitochondria in the heart is essential, and drug-induced murine models causing mitochondrial damage result in cardiovascular arrhythmias and cardiomyopathy [112]. Humans are not the only species to have mitochondrial gene mutations associated with CM. Mutations in *PDK4*, a mitochondrial gene, are associated with Doberman pinscher DCM in the dog [58], therefore, it is essential that new studies also investigate mitochondrial genes.

Peri- or postpartum cardiomyopathy (PPCM) has been described under a number of conditions, but is defined by the European Society of Cardiology as the “development of heart failure toward the end of pregnancy or in the months following delivery” [113]. This has been observed in many species including human, canine, and bovine cases [113–115]. Much of the literature indicates that peripartum cardiomyopathy might be better referred to as DCM that is initiated during pregnancy or soon thereafter, but discussions are still ongoing as to whether this is an accurate portrayal [116]. In the case of peripartum CM, it has been shown that oxidative stress and prolactin play roles in the disease in humans and mouse models [116,117]. Of particular interest are the few human genetic studies that have been carried out to date, all of which associate PPCM with DCM causing mutations in *TNNC1* and *TTN* in humans [118–120]. The complex nature of this cardiomyopathy, potential overlaps and links to DCM, and the frequently observed hypertension, preeclampsia, and altered hormone levels make this a difficult CM to investigate, and more work needs to be undertaken, not only in humans but in other species too.

7. Conclusions

Common genetic pathways could exist among cardiomyopathies and among species. As discussed above, there are multiple genes where differing mutations within each gene can cause different CMs in humans. Equally, there are many examples where genes have been implied in humans but not non-human animals—and vice versa—such as the striatin gene mutations, which are associated with both ARVC and DCM in Boxer dogs [57]. It is also clear that different mutations in the same gene can cause different types of CM. Both DCM and HCM in humans have been linked to *TTN*, *MYH6*, *MYH7*, *MYBPC3*, *TNNT2*, *TNNI3*, *TPM1*, *ACTC/ACTC1*, *TNNC1*, *ACTN2*, *ANKDR1*, *CSRFP3*, *LDB3*, *TCAP*, *VCL*, *PLN*, and *RYR2* [17,52]; likewise, mutations in *MYH7* and *TNNI3* can cause RCM, DCM, and HCM [40,121]. *RYR2* has been implicated in ARVC, DCM, and HCM [86,121], and the ARVC genes

DSP, *PKP2*, *DSG2*, *DSC2*, and *JUP* have all been associated to DCM [17]. Upon tabulation of 50 genes causing HCM, DCM, and ARVC, it was shown that the degree of heterogeneity in contributing genes is considerable [18]. In their review of 50 genes causing HCM, DCM, and ARVC, only 7 genes had changes unique to HCM and 15 genes had changes unique to DCM, while changes in 33 other genes contributed to both disorders. Similarly, ARVC currently has 3 genes contributing solely to ARVC and a further 7 genes associated with DCM, HCM, and ARVC. Most of the mutations causing ARVC are found in the genes *PKP2* and in the desmosomal cadherins, and these genes are also associated with DCM. Many of the CM-associated mutations are present in the structural genes, which are phylogenetically highly conserved; therefore, the frequent similarities in mutations/genes affected between humans and animals are not surprising. This further supports the use of multiple species investigations when looking at the differing genotypes in order to understand the cardiomyopathies. A summary of all genes associated with naturally occurring CMs in each species has been compiled in Table A1.

With both genotype and phenotype overlapping greatly between the different cardiomyopathies, it is likely that, in the future, diagnosis will rely greatly on next-generation sequencing (NGS) technology and genome-wide association studies. Lessons may be learned about the genetic causes of CMs using information from these studies, also. Many of the studies, both human and animal, have previously relied on familial studies or cohorts with relatively small numbers. Over the years, study sizes have increased. As more advanced technology is more readily available, and at a lower cost, the candidate gene approach is being replaced with larger sequencing studies on larger cohorts. Examples of this are already observed throughout the literature in this review, but national and international endeavours such as the 100,000 genome project [122] are utilising the power of mutation and disorder detection, not only in common disorders but also in rare CMs. Large-scale studies such as these will frequently have to be supported by cohort studies. A number of papers have suggested that mutations in specific genes are not exclusively involved with particular CMs in individual species or breeds or animals, rather that different mutations in the same genes could well cause differing CMs [123–125]. Critical analysis of the sample sizes should be carried out before genes and/or mutations are ruled out. The genetic heterogeneity of CM genes can be indicated with the knowledge that a recent diagnostic NGS panel for CM diagnosis has 104 genes and candidate genes designed from research papers in the field [126]. Testing not only humans, but animals too, will not only aid in diagnosis and prognosis, but potentially assist with understanding epidemiology of the disorder. Genetic testing will also assist with healthcare options and treatment plans even prior to clinical symptoms of the CM, and aid in the reduction of affected animals within breeds. Although many of the cardiomyopathies characterised to date are single gene disorders, there is increasing evidence that multiple gene associations can contribute towards this disease. This has been evidenced in dogs [124,125], but more research needs to be undertaken in order to understand the situation for each CM type in each species.

Cardiomyopathies are complex cardiovascular disorders, but advances in genetic detection are important not only to humans but also in animals, as models of the human condition, but also in order to advance non-human healthcare and breeding programmes. Targeted healthcare, diagnosis and prognosis are essential for cardiomyopathy patients, and further insights into the genetic causes are essential.

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Table A1. Cont.

Gene	DCM				HCM		ARVC			RCM	References
	Human	Canine	Bovine	Turkey	Human	Feline	Human	Canine	Bovine	Human	
<i>FKTN</i> Fukutin	Y										[162]
<i>FLNC</i> Filamin C										Y	[75]
<i>FOXD4</i> Forkhead Box D4	Y										[164]
<i>FSTL5</i> Follistatin Like 5		Y									[58]
<i>GATAD1</i> GATA Zinc Finger Domain Containing 1	Y										[165]
<i>HAND1</i> Heart and Neural Crest Derivatives Expressed 1	Y										[166]
<i>HCG22</i> HLA Complex Group 22	Y										[167]
<i>HLA-DQB1</i> Major Histocompatibility Complex, Class II, DQ Beta 1	Y										[167]
<i>HSPB7</i> Heat Shock Protein Family B (Small) Member 7	Y										[168]
<i>ILK</i> Integrin Linked Kinase	Y										[169]
<i>JPH2</i> Junctophilin 2						Y					[170]
<i>JUP</i> Junction Plakoglobin								Y			[171]
<i>LAMA2</i> Laminin Subunit Alpha 2	Y										[172]
<i>LAMA4</i> Laminin Subunit Alpha 4	Y										[169]
<i>LAMP2</i> Lysosomal Associated Membrane Protein 2	Y									Y	[75,173]

References

1. Mathers, C.D.; Loncar, D. Projections of global mortality and burden of disease from 2002 to 2030. *PLoS Med.* **2006**, *3*, e442. [[CrossRef](#)] [[PubMed](#)]
2. Cheng, Y.; Hogarth, K.A.; O'Sullivan, M.L.; Regnier, M.; Pyle, W.G. 2-deoxyadenosine triphosphate restores the contractile function of cardiac myofibril from adult dogs with naturally occurring dilated cardiomyopathy. *Am. J. Physiol. Heart Circ. Physiol.* **2016**, *310*, H80–H91. [[CrossRef](#)] [[PubMed](#)]
3. Houser, S.R.; Margulies, K.B.; Murphy, A.M.; Spinale, F.G.; Francis, G.S.; Prabhu, S.D.; Rockman, H.A.; Kass, D.A.; Molkentin, J.D.; Sussman, M.A.; et al. Animal models of heart failure: A scientific statement from the american heart association. *Circ. Res.* **2012**, *111*, 131–150. [[CrossRef](#)] [[PubMed](#)]
4. Elliott, P.; Andersson, B.; Arbustini, E.; Bilinska, Z.; Cecchi, F.; Charron, P.; Dubourg, O.; Kuhl, U.; Maisch, B.; McKenna, W.J.; et al. Classification of the cardiomyopathies: A position statement from the european society of cardiology working group on myocardial and pericardial diseases. *Eur. Heart J.* **2008**, *29*, 270–276. [[CrossRef](#)] [[PubMed](#)]
5. Maron, B.J.; Towbin, J.A.; Thiene, G.; Antzelevitch, C.; Corrado, D.; Arnett, D.; Moss, A.J.; Seidman, C.E.; Young, J.B. Contemporary definitions and classification of the cardiomyopathies: An american heart association scientific statement from the council on clinical cardiology, heart failure and transplantation committee; quality of care and outcomes research and functional genomics and translational biology interdisciplinary working groups; and council on epidemiology and prevention. *Circulation* **2006**, *113*, 1807–1816. [[PubMed](#)]
6. Strong, V.J.; Grindlay, D.; Redrobe, S.; Cobb, M.; White, K. A systematic review of the literature relating to captive great ape morbidity and mortality. *J. Zoo Wildl. Med.* **2016**, *47*, 697–710. [[CrossRef](#)] [[PubMed](#)]
7. Twycross. Ape heart project. Available online: <https://twycrosszoo.org/conservation/research-at-twycross-zoo/current-research/ape-heart-project/> (accessed on 28 December 2016).
8. Fatkin, D. Guidelines for the diagnosis and management of familial dilated cardiomyopathy. *Heart Lung Circ.* **2007**, *16*, 19–21. [[CrossRef](#)] [[PubMed](#)]
9. Herman, D.S.; Lam, L.; Taylor, M.R.; Wang, L.; Teekakirikul, P.; Christodoulou, D.; Conner, L.; DePalma, S.R.; McDonough, B.; Sparks, E.; et al. Truncations of titin causing dilated cardiomyopathy. *N. Engl. J. Med.* **2012**, *366*, 619–628. [[CrossRef](#)] [[PubMed](#)]
10. Nihoyannopoulos, P.; Dawson, D. Restrictive cardiomyopathies. *Eur. J. Echocardiogr.* **2009**, *10*, iii23–iii33. [[CrossRef](#)] [[PubMed](#)]
11. Garcia-Castro, M.; Reguero, J.R.; Alvarez, V.; Batalla, A.; Soto, M.I.; Albaladejo, V.; Coto, E. Hypertrophic cardiomyopathy linked to homozygosity for a new mutation in the myosin-binding protein c gene (a627v) suggests a dosage effect. *Int. J. Cardiol.* **2005**, *102*, 501–507. [[CrossRef](#)] [[PubMed](#)]
12. Hoedemaekers, Y.M.; Caliskan, K.; Majoor-Krakauer, D.; van de Laar, I.; Michels, M.; Witsenburg, M.; ten Cate, F.J.; Simoons, M.L.; Dooijes, D. Cardiac beta-myosin heavy chain defects in two families with non-compaction cardiomyopathy: Linking non-compaction to hypertrophic, restrictive, and dilated cardiomyopathies. *Eur. Heart J.* **2007**, *28*, 2732–2737. [[CrossRef](#)] [[PubMed](#)]
13. Jarcho, J.A.; McKenna, W.; Pare, J.A.; Solomon, S.D.; Holcombe, R.F.; Dickie, S.; Levi, T.; Donis-Keller, H.; Seidman, J.G.; Seidman, C.E. Mapping a gene for familial hypertrophic cardiomyopathy to chromosome 14q1. *N. Engl. J. Med.* **1989**, *321*, 1372–1378. [[CrossRef](#)] [[PubMed](#)]
14. Geisterfer-Lowrance, A.A.; Kass, S.; Tanigawa, G.; Vosberg, H.P.; McKenna, W.; Seidman, C.E.; Seidman, J.G. A molecular basis for familial hypertrophic cardiomyopathy: A beta cardiac myosin heavy chain gene missense mutation. *Cell* **1990**, *62*, 999–1006. [[CrossRef](#)]
15. Elliott, P.M.; Anastakis, A.; Borger, M.A.; Borggrefe, M.; Cecchi, F.; Charron, P.; Hagege, A.A.; Lafont, A.; Limongelli, G.; Mahrholdt, H.; et al. 2014 ESC Guidelines on diagnosis and management of hypertrophic cardiomyopathy. The Task Force for the Diagnosis and Management of Hypertrophic Cardiomyopathy of the European Society of Cardiology (ESC). *Eur. Heart J.* **2014**, *35*, 2733–2779. [[PubMed](#)]
16. Maron, B.J.; Maron, M.S.; Semsarian, C. Genetics of hypertrophic cardiomyopathy after 20 years: Clinical perspectives. *J. Am. Coll. Cardiol.* **2012**, *60*, 705–715. [[CrossRef](#)] [[PubMed](#)]
17. Landstrom, A.P.; Ackerman, M.J. Mutation type is not clinically useful in predicting prognosis in hypertrophic cardiomyopathy. *Circulation* **2010**, *122*, 2441–2449; discussion 2450. [[CrossRef](#)] [[PubMed](#)]

18. McNally, E.M.; Golbus, J.R.; Puckelwartz, M.J. Genetic mutations and mechanisms in dilated cardiomyopathy. *J. Clin. Investig.* **2013**, *123*, 19–26. [[CrossRef](#)] [[PubMed](#)]
19. Fox, P.R.; Liu, S.-K.; Maron, B.J. Echocardiographic assessment of spontaneously occurring feline hypertrophic cardiomyopathy. An Animal Model of Human Disease. *Circulation* **1995**, *92*, 2645–2651. [[PubMed](#)]
20. Egenvall, A.; Bonnett, B.N.; Häggström, J. Heart disease as a cause of death in insured swedish dogs younger than 10 years of age. *J. Vet. Intern. Med.* **2006**, *20*, 894–903. [[CrossRef](#)] [[PubMed](#)]
21. Washizu, M.; Takemura, N.; Machida, N.; Nawa, H.; Yamamoto, T.; Mitake, H.; Washizu, T. Hypertrophic cardiomyopathy in an aged dog. *J. Vet. Med. Sci.* **2003**, *65*, 753–756. [[CrossRef](#)] [[PubMed](#)]
22. Paige, C.F.; Abbott, J.A.; Elvinger, F.; Pyle, R.L. Prevalence of cardiomyopathy in apparently healthy cats. *J. Am. Vet. Med. Assoc.* **2009**, *234*, 1398–1403. [[CrossRef](#)] [[PubMed](#)]
23. Kittleson, M.D.; Meurs, K.M.; Munro, M.J.; Kittleson, J.A.; Liu, S.K.; Pion, P.D.; Towbin, J.A. Familial hypertrophic cardiomyopathy in maine coon cats: An animal model of human disease. *Circulation* **1999**, *99*, 3172–3180. [[CrossRef](#)] [[PubMed](#)]
24. Meurs, K.M.; Norgard, M.M.; Ederer, M.M.; Hendrix, K.P.; Kittleson, M.D. A substitution mutation in the myosin binding protein c gene in ragdoll hypertrophic cardiomyopathy. *Genomics* **2007**, *90*, 261–264. [[CrossRef](#)] [[PubMed](#)]
25. Langhorn, R.; Willesen, J.L.; Tarnow, I.; Kjelgaard-Hansen, M.; Koch, J. Cardiac troponin i in three cat breeds with hypertrophic cardiomyopathy. *Vet. Record* **2016**, *178*, 532. [[CrossRef](#)] [[PubMed](#)]
26. Meurs, K.M.; Sanchez, X.; David, R.M.; Bowles, N.E.; Towbin, J.A.; Reiser, P.J.; Kittleson, J.A.; Munro, M.J.; Dryburgh, K.; Macdonald, K.A.; et al. A cardiac myosin binding protein c mutation in the maine coon cat with familial hypertrophic cardiomyopathy. *Hum. Mol. Genet.* **2005**, *14*, 3587–3593. [[CrossRef](#)] [[PubMed](#)]
27. Wess, G.; Schinner, C.; Weber, K.; Kuchenhoff, H.; Hartmann, K. Association of a31p and a74t polymorphisms in the myosin binding protein c3 gene and hypertrophic cardiomyopathy in maine coon and other breed cats. *J. Vet. Intern. Med.* **2010**, *24*, 527–532. [[CrossRef](#)] [[PubMed](#)]
28. Watkins, H.; Conner, D.; Thierfelder, L.; Jarcho, J.A.; MacRae, C.; McKenna, W.J.; Maron, B.J.; Seidman, J.G.; Seidman, C.E. Mutations in the cardiac myosin binding protein-c gene on chromosome 11 cause familial hypertrophic cardiomyopathy. *Nat. Genet.* **1995**, *11*, 434–437. [[CrossRef](#)] [[PubMed](#)]
29. Borgeat, K.; Sherwood, K.; Payne, J.R.; Luis Fuentes, V.; Connolly, D.J. Plasma cardiac troponin i concentration and cardiac death in cats with hypertrophic cardiomyopathy. *J. Vet. Intern. Med.* **2014**, *28*, 1731–1737. [[CrossRef](#)] [[PubMed](#)]
30. Huang, S.Y.; Tsou, H.L.; Chiu, Y.T.; Shyu, J.J.; Wu, J.J.; Lin, J.H.; Liu, S.K. Heritability estimate of hypertrophic cardiomyopathy in pigs (*sus scrofa domestica*). *Lab. Anim. Sci.* **1996**, *46*, 310–314. [[PubMed](#)]
31. Tardiff, J.C.; Hewett, T.E.; Palmer, B.M.; Olsson, C.; Factor, S.M.; Moore, R.L.; Robbins, J.; Leinwand, L.A. Cardiac troponin t mutations result in allele-specific phenotypes in a mouse model for hypertrophic cardiomyopathy. *J. Clin. Investig.* **1999**, *104*, 469–481. [[CrossRef](#)] [[PubMed](#)]
32. Marian, A.J.; Wu, Y.; Lim, D.-S.; McCluggage, M.; Youker, K.; Yu, Q.-t.; Brugada, R.; DeMayo, F.; Quinones, M.; Roberts, R. A transgenic rabbit model for human hypertrophic cardiomyopathy. *J. Clin. Investig.* **1999**, *104*, 1683–1692. [[CrossRef](#)] [[PubMed](#)]
33. Maekawa, K.; Hirayama, A.; Iwata, Y.; Tajima, Y.; Nishimaki-Mogami, T.; Sugawara, S.; Ueno, N.; Abe, H.; Ishikawa, M.; Murayama, M.; et al. Global metabolomic analysis of heart tissue in a hamster model for dilated cardiomyopathy. *J. Mol. Cell. Cardiol.* **2013**, *59*, 76–85.
34. Sakamoto, A. Molecular pathogenesis of severe cardiomyopathy in the to-2 hamster. *Exp. Clin. Cardiol.* **2003**, *8*, 143–146. [[PubMed](#)]
35. Sakamoto, A.; Ono, K.; Abe, M.; Jasmin, G.; Eki, T.; Murakami, Y.; Masaki, T.; Toyooka, T.; Hanaoka, F. Both hypertrophic and dilated cardiomyopathies are caused by mutation of the same gene, delta-sarcoglycan, in hamster: An animal model of disrupted dystrophin-associated glycoprotein complex. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 13873–13878. [[CrossRef](#)] [[PubMed](#)]
36. Guttman, O.P.; Mohiddin, S.A.; Elliott, P.M. Almanac 2014: Cardiomyopathies. *Heart* **2014**, *100*, 756–764. [[CrossRef](#)] [[PubMed](#)]
37. Raju, H.; Alberg, C.; Sagoo, G.S.; Burton, H.; Behr, E.R. Inherited cardiomyopathies. *Br. Med. J.* **2011**, *343*, d6966. [[CrossRef](#)] [[PubMed](#)]

38. Hershberger, R.E.; Morales, A.; Siegfried, J.D. Clinical and genetic issues in dilated cardiomyopathy: A review for genetics professionals. *Genet. Med.* **2010**, *12*, 655–667. [[CrossRef](#)] [[PubMed](#)]
39. Codd, M.B.; Sugrue, D.D.; Gersh, B.J.; Melton, L.J. Epidemiology of idiopathic dilated and hypertrophic cardiomyopathy. A population-based study in olmsted county, minnesota, 1975–1984. *Circulation* **1989**, *80*, 564–572. [[CrossRef](#)] [[PubMed](#)]
40. Posafalvi, A.; Herkert, J.C.; Sinke, R.J.; van den Berg, M.P.; Mogensen, J.; Jongbloed, J.D.; van Tintelen, J.P. Clinical utility gene card for: Dilated cardiomyopathy (cmd). *Eur. J. Hum. Genet.* **2013**, *21*. [[CrossRef](#)] [[PubMed](#)]
41. Dukes-McEwan, J.; Borgarelli, M.; Tidholm, A.; Vollmar, A.C.; Haggstrom, J. Proposed guidelines for the diagnosis of canine idiopathic dilated cardiomyopathy. *J. Vet. Cardiol.* **2003**, *5*, 7–19. [[CrossRef](#)]
42. Hambrook, L.E.; Bennett, P.F. Effect of pimobendan on the clinical outcome and survival of cats with non-aurine responsive dilated cardiomyopathy. *J. Feline Med. Surg.* **2012**, *14*, 233–239. [[CrossRef](#)] [[PubMed](#)]
43. Weekes, J.; Wheeler, C.H.; Yan, J.X.; Weil, J.; Eschenhagen, T.; Scholtysik, G.; Dunn, M.J. Bovine dilated cardiomyopathy: Proteomic analysis of an animal model of human dilated cardiomyopathy. *Electrophoresis* **1999**, *20*, 898–906. [[CrossRef](#)]
44. Owczarek-Lipska, M.; Plattet, P.; Zipperle, L.; Drogemuller, C.; Posthaus, H.; Dolf, G.; Braunschweig, M.H. A nonsense mutation in the optic atrophy 3 gene (opa3) causes dilated cardiomyopathy in red holstein cattle. *Genomics* **2011**, *97*, 51–57. [[CrossRef](#)] [[PubMed](#)]
45. Nart, P.; Thompson, H.; Barrett, D.C.; Armstrong, S.C.; McPhaden, A.R. Clinical and pathological features of dilated cardiomyopathy in holstein-friesian cattle. *Vet. Record* **2004**, *155*, 355–361. [[CrossRef](#)]
46. Biesiadecki, B.J.; Jin, J.-P. Exon skipping in cardiac troponin t of turkeys with inherited dilated cardiomyopathy. *J. Biol. Chem.* **2002**, *277*, 18459–18468. [[CrossRef](#)] [[PubMed](#)]
47. Frame, D.D.; Kelly, E.J.; Van Wettere, A. Dilated cardiomyopathy in a rio grande wild turkey (meleagris gallopavo intermedia) in southern Utah, USA, 2013. *J. Wildl. Dis.* **2015**, *51*, 790–792. [[CrossRef](#)] [[PubMed](#)]
48. Wilson, F.D.; Magee, D.L.; Jones, K.H.; Baravik-Munsell, E.; Cummings, T.S.; Wills, R.W.; Pace, L.W. Morphometric documentation of a high prevalence of left ventricular dilated cardiomyopathy in both clinically normal and cyanotic mature commercial broiler breeder roosters with comparisons to market-age broilers. *Avian Dis.* **2016**, *60*, 589–595. [[CrossRef](#)] [[PubMed](#)]
49. Pion, P.D.; Kittleson, M.D.; Rogers, Q.R.; Morris, J.G. Myocardial failure in cats associated with low plasma taurine: A reversible cardiomyopathy. *Science* **1987**, *237*, 764–768. [[CrossRef](#)] [[PubMed](#)]
50. Ferasin, L.; Sturgess, C.P.; Cannon, M.J.; Caney, S.M.; Gruffydd-Jones, T.J.; Wotton, P.R. Feline idiopathic cardiomyopathy: A retrospective study of 106 cats (1994–2001). *J. Feline Med. Surg.* **2003**, *5*, 151–159. [[CrossRef](#)]
51. Lawler, D.F.; Templeton, A.J.; Monti, K.L. Evidence for genetic involvement in feline dilated cardiomyopathy. *J. Vet. Intern. Med.* **1993**, *7*, 383–387. [[CrossRef](#)] [[PubMed](#)]
52. Simpson, S.; Edwards, J.; Ferguson-Mignan, T.F.N.; Cobb, M.; Mongan, N.P.; Rutland, C.S. Genetics of human and canine dilated cardiomyopathy. *Int. J. Genom.* **2015**. [[CrossRef](#)] [[PubMed](#)]
53. Towbin, J.A.; Lowe, A.M.; Colan, S.D.; Sleeper, L.A.; Orav, E.J.; Clunie, S.; Messere, J.; Cox, G.F.; Lurie, P.R.; Hsu, D.; et al. Incidence, causes, and outcomes of dilated cardiomyopathy in children. *J. Am. Med. Assoc.* **2006**, *296*, 1867–1876. [[CrossRef](#)] [[PubMed](#)]
54. Dambach, D.M.; Lannon, A.; Sleeper, M.M.; Buchanan, J. Familial dilated cardiomyopathy of young portuguese water dogs. *J. Vet. Intern. Med.* **1999**, *13*, 65–71. [[CrossRef](#)] [[PubMed](#)]
55. Mausberg, T.B.; Wess, G.; Simak, J.; Keller, L.; Drogemuller, M.; Drogemuller, C.; Webster, M.T.; Stephenson, H.; Dukes-McEwan, J.; Leeb, T. A locus on chromosome 5 is associated with dilated cardiomyopathy in doberman pinschers. *PLoS ONE* **2011**, *6*. [[CrossRef](#)] [[PubMed](#)]
56. Meurs, K.M.; Lahmers, S.; Keene, B.W.; White, S.N.; Oyama, M.A.; Mauceli, E.; Lindblad-Toh, K. A splice site mutation in a gene encoding for pdk4, a mitochondrial protein, is associated with the development of dilated cardiomyopathy in the Doberman pinscher. *Hum. Genet.* **2012**, *131*, 1319–1325. [[CrossRef](#)] [[PubMed](#)]
57. Meurs, K.M.; Stern, J.A.; Sisson, D.D.; Kittleson, M.D.; Cunningham, S.M.; Ames, M.K.; Atkins, C.E.; DeFrancesco, T.; Hodge, T.E.; Keene, B.W.; et al. Association of dilated cardiomyopathy with the striatin mutation genotype in boxer dogs. *J. Vet. Intern. Med.* **2013**, *27*, 1437–1440. [[PubMed](#)]
58. Philipp, U.; Vollmar, A.; Haggstrom, J.; Thomas, A.; Distl, O. Multiple loci are associated with dilated cardiomyopathy in Irish wolfhounds. *PLoS ONE* **2012**, *7*. [[CrossRef](#)] [[PubMed](#)]

59. Schatzberg, S.J.; Olby, N.J.; Breen, M.; Anderson, L.V.B.; Langford, C.F.; Dickens, H.F.; Wilton, S.D.; Zeiss, C.J.; Binns, M.M.; Kornegay, J.N.; et al. Molecular analysis of a spontaneous dystrophin ‘knockout’ dog. *Neuromuscular Disord.* **1999**, *9*, 289–295. [[CrossRef](#)]
60. Werner, P.; Raducha, M.G.; Prociuk, U.; Sleeper, M.M.; Van Winkle, T.J.; Henthorn, P.S. A novel locus for dilated cardiomyopathy maps to canine chromosome 8. *Genomics* **2008**, *91*, 517–521. [[CrossRef](#)] [[PubMed](#)]
61. Dolf, G.; Stricker, C.; Tontis, A.; Martig, J.; Gaillard, C. Evidence for autosomal recessive inheritance of a major gene for bovine dilated cardiomyopathy. *J. Anim. Sci.* **1998**, *76*, 1824–1829. [[CrossRef](#)] [[PubMed](#)]
62. Pinto, J.R.; Yang, S.W.; Hitz, M.P.; Parvatiyar, M.S.; Jones, M.A.; Liang, J.; Kokta, V.; Talajic, M.; Tremblay, N.; Jaeggi, M.; et al. Fetal cardiac troponin isoforms rescue the increased ca²⁺ sensitivity produced by a novel double deletion in cardiac troponin t linked to restrictive cardiomyopathy: A clinical, genetic, and functional approach. *J. Biol. Chem.* **2011**, *286*, 20901–20912. [[CrossRef](#)] [[PubMed](#)]
63. Manning, E.P.; Guinto, P.J.; Tardiff, J.C. Correlation of molecular and functional effects of mutations in cardiac troponin t linked to familial hypertrophic cardiomyopathy: An integrative in silico/in vitro approach. *J. Biol. Chem.* **2012**, *287*, 14515–14523. [[CrossRef](#)] [[PubMed](#)]
64. Rutland, C.S.; Polo-Parada, L.; Ehler, E.; Alibhai, A.; Thorpe, A.; Suren, S.; Emes, R.D.; Patel, B.; Loughna, S. Knockdown of embryonic myosin heavy chain reveals an essential role in the morphology and function of the developing heart. *Development* **2011**, *138*, 3955–3966. [[CrossRef](#)] [[PubMed](#)]
65. Rutland, C.; Warner, L.; Thorpe, A.; Alibhai, A.; Robinson, T.; Shaw, B.; Layfield, R.; Brook, J.D.; Loughna, S. Knockdown of alpha myosin heavy chain disrupts the cytoskeleton and leads to multiple defects during chick cardiogenesis. *J. Anat.* **2009**, *214*, 905–915. [[CrossRef](#)] [[PubMed](#)]
66. Granados-Riveron, J.T.; Ghosh, T.K.; Pope, M.; Bu’Lock, F.; Thornborough, C.; Eason, J.; Kirk, E.P.; Fatkin, D.; Feneley, M.P.; Harvey, R.P.; et al. Alpha-cardiac myosin heavy chain (myh6) mutations affecting myofibril formation are associated with congenital heart defects. *Hum. Mol. Genet.* **2010**, *19*, 4007–4016. [[CrossRef](#)] [[PubMed](#)]
67. Carniel, E.; Taylor, M.R.; Sinagra, G.; Di Lenarda, A.; Ku, L.; Fain, P.R.; Boucek, M.M.; Cavanaugh, J.; Miodic, S.; Slavov, D.; et al. Alpha-myosin heavy chain: A sarcomeric gene associated with dilated and hypertrophic phenotypes of cardiomyopathy. *Circulation* **2005**, *112*, 54–59. [[CrossRef](#)] [[PubMed](#)]
68. Toyoda, Y.; Okada, M.; Kashem, M.A. A canine model of dilated cardiomyopathy induced by repetitive intracoronary doxorubicin administration. *J. Thorac. Cardiovasc. Surg.* **1998**, *115*, 1367–1373. [[CrossRef](#)]
69. Christiansen, S.; Redmann, K.; Scheld, H.H.; Jahn, U.R.; Stypmann, J.; Fobker, M.; Gruber, A.D.; Hammel, D. Adriamycin-induced cardiomyopathy in the dog—An appropriate model for research on partial left ventriculectomy? *J. Heart Lung Transplant.* **2002**, *21*, 783–790. [[CrossRef](#)]
70. Nikolaidis, L.A.; Elahi, D.; Hentosz, T.; Doverspike, A.; Huerbin, R.; Zourelis, L.; Stolarski, C.; Shen, Y.T.; Shannon, R.P. Recombinant glucagon-like peptide-1 increases myocardial glucose uptake and improves left ventricular performance in conscious dogs with pacing-induced dilated cardiomyopathy. *Circulation* **2004**, *110*, 955–961. [[CrossRef](#)] [[PubMed](#)]
71. Brodehl, A.; Ferrier, R.A.; Hamilton, S.J.; Greenway, S.C.; Brundler, M.-A.; Yu, W.; Gibson, W.T.; McKinnon, M.L.; McGillivray, B.; Alvarez, N.; et al. Mutations in flnc are associated with familial restrictive cardiomyopathy. *Hum. Mutat.* **2016**, *37*, 269–279. [[CrossRef](#)] [[PubMed](#)]
72. Sasaki, N.; Garcia, M.; Ko, H.H.; Sharma, S.; Parness, I.A.; Srivastava, S. Applicability of published guidelines for assessment of left ventricular diastolic function in adults to children with restrictive cardiomyopathy: An observational study. *Pediatr. Cardiol.* **2015**, *36*, 386–392. [[CrossRef](#)] [[PubMed](#)]
73. Kubo, T.; Gimeno, J.R.; Bahl, A.; Steffensen, U.; Steffensen, M.; Osman, E.; Thaman, R.; Mogensen, J.; Elliott, P.M.; Doi, Y.; et al. Prevalence, clinical significance, and genetic basis of hypertrophic cardiomyopathy with restrictive phenotype. *J. Am. Coll. Cardiol.* **2007**, *49*, 2419–2426. [[CrossRef](#)] [[PubMed](#)]
74. Mogensen, J.; Kubo, T.; Duque, M.; Uribe, W.; Shaw, A.; Murphy, R.; Gimeno, J.R.; Elliott, P.; McKenna, W.J. Idiopathic restrictive cardiomyopathy is part of the clinical expression of cardiac troponin i mutations. *J. Clin. Investig.* **2003**, *111*, 209–216. [[CrossRef](#)] [[PubMed](#)]
75. Gallego-Delgado, M.; Delgado, J.F.; Brossa-Loidi, V.; Palomo, J.; Marzoa-Rivas, R.; Perez-Villa, F.; Salazar-Mendiguchia, J.; Ruiz-Cano, M.J.; Gonzalez-Lopez, E.; Padron-Barthe, L.; et al. Idiopathic restrictive cardiomyopathy is primarily a genetic disease. *J. Am. Coll. Cardiol.* **2016**, *67*, 3021–3023. [[PubMed](#)]

76. Mouton, J.M.; Pellizzon, A.S.; Goosen, A.; Kinnear, C.J.; Herbst, P.G.; Brink, P.A.; Moolman-Smook, J.C. Diagnostic disparity and identification of two *tnni3* gene mutations, one novel and one arising de novo, in south african patients with restrictive cardiomyopathy and focal ventricular hypertrophy. *Cardiovasc. J. Afr.* **2015**, *26*, 63–69. [[CrossRef](#)] [[PubMed](#)]
77. Fox, P.R.; Basso, C.; Thiene, G.; Maron, B.J. Spontaneously occurring restrictive nonhypertrophied cardiomyopathy in domestic cats: A new animal model of human disease. *Cardiovasc. Pathol.* **2014**, *23*, 28–34. [[CrossRef](#)] [[PubMed](#)]
78. Kimura, Y.; Karakama, S.; Hirakawa, A.; Tsuchiaka, S.; Kobayashi, M.; Machida, N. Pathological features and pathogenesis of the endomyocardial form of restrictive cardiomyopathy in cats. *J. Comp. Pathol.* **2016**, *155*, 190–198. [[CrossRef](#)] [[PubMed](#)]
79. Davis, J.; Yasuda, S.; Palpant, N.J.; Martindale, J.; Stevenson, T.; Converso, K.; Metzger, J.M. Diastolic dysfunction and thin filament dysregulation resulting from excitation-contraction uncoupling in a mouse model of restrictive cardiomyopathy. *J. Mol. Cell. Cardiol.* **2012**, *53*, 446–457. [[CrossRef](#)] [[PubMed](#)]
80. Bakeer, N.; James, J.; Roy, S.; Wansapura, J.; Shanmukhappa, S.K.; Lorenz, J.N.; Osinska, H.; Backer, K.; Huby, A.C.; Shrestha, A.; et al. Sick cell anemia mice develop a unique cardiomyopathy with restrictive physiology. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E5182–5191. [[CrossRef](#)] [[PubMed](#)]
81. Niss, O.; Quinn, C.T.; Lane, A.; Daily, J.; Khoury, P.R.; Bakeer, N.; Kimball, T.R.; Towbin, J.A.; Malik, P.; Taylor, M.D. Cardiomyopathy with restrictive physiology in sickle cell disease. *JACC Cardiovasc. Imaging* **2016**, *9*, 243–252.
82. Basso, C.; Corrado, D.; Marcus, F.I.; Nava, A.; Thiene, G. Arrhythmogenic right ventricular cardiomyopathy. *Lancet* **2009**, *373*, 1289–1300. [[CrossRef](#)]
83. Ruwald, A.-C.; Marcus, F.; Estes, N.A.M.; Link, M.; McNitt, S.; Polonsky, B.; Calkins, H.; Towbin, J.A.; Moss, A.J.; Zareba, W. Association of competitive and recreational sport participation with cardiac events in patients with arrhythmogenic right ventricular cardiomyopathy: Results from the north american multidisciplinary study of arrhythmogenic right ventricular cardiomyopathy. *Eur. Heart J.* **2015**, *36*, 1735–1743. [[CrossRef](#)] [[PubMed](#)]
84. Elliott, P.; O'Mahony, C.; Syrris, P.; Evans, A.; Rivera Sorensen, C.; Sheppard, M.N.; Carr-White, G.; Pantazis, A.; McKenna, W.J. Prevalence of desmosomal protein gene mutations in patients with dilated cardiomyopathy. *Circ. Cardiovasc. Genet.* **2010**, *3*, 314–322. [[CrossRef](#)] [[PubMed](#)]
85. Lazzarini, E.; Jongbloed, J.D.; Pilichou, K.; Thiene, G.; Basso, C.; Bikker, H.; Charbon, B.; Swertz, M.; van Tintelen, J.P.; van der Zwaag, P.A. The *arvd/c* genetic variants database: 2014 update. *Hum. Mutat.* **2015**, *36*, 403–410. [[CrossRef](#)] [[PubMed](#)]
86. Patel, H.; Shah, P.; Rampal, U.; Shamoan, F.; Tiyyagura, S. Arrhythmogenic right ventricular dysplasia/cardiomyopathy (*arvd/c*) and catecholaminergic polymorphic ventricular tachycardia (*cpvt*): A phenotypic spectrum seen in same patient. *J. Electrocardiol.* **2015**, *48*, 874–878. [[CrossRef](#)] [[PubMed](#)]
87. Kucerova, D.; Doka, G.; Kruzliak, P.; Turcekova, K.; Kmecova, J.; Brnoliakova, Z.; Kyselovic, J.; Kirchhefer, U.; Muller, F.U.; Krenek, P.; et al. Unbalanced upregulation of ryanodine receptor 2 plays a particular role in early development of daunorubicin cardiomyopathy. *Am. J. Transl. Res.* **2015**, *7*, 1280–1294. [[PubMed](#)]
88. Severini, G.M.; Krajcinovic, M.; Pinamonti, B.; Sinagra, G.; Fioretti, P.; Brunazzi, M.C.; Falaschi, A.; Camerini, F.; Giacca, M.; Mestroni, L. A new locus for arrhythmogenic right ventricular dysplasia on the long arm of chromosome 14. *Genomics* **1996**, *31*, 193–200. [[CrossRef](#)] [[PubMed](#)]
89. Rampazzo, A.; Nava, A.; Miorin, M.; Fonderico, P.; Pope, B.; Tiso, N.; Livolsi, B.; Zimbello, R.; Thiene, G.; Danieli, G.A. *Arvd4*, a new locus for arrhythmogenic right ventricular cardiomyopathy, maps to chromosome 2 long arm. *Genomics* **1997**, *45*, 259–263. [[CrossRef](#)] [[PubMed](#)]
90. Li, D.; Ahmad, F.; Gardner, M.J.; Weillbaeher, D.; Hill, R.; Karibe, A.; Gonzalez, O.; Tapscott, T.; Sharratt, G.P.; Bachinski, L.L.; et al. The locus of a novel gene responsible for arrhythmogenic right-ventricular dysplasia characterized by early onset and high penetrance maps to chromosome 10p12-p14. *Am. J. Hum. Genet.* **2000**, *66*, 148–156. [[CrossRef](#)] [[PubMed](#)]
91. Matolwani, L.O.; Bardien, S.; Rebello, G.; Oppon, E.; Munclinger, M.; Ramesar, R.; Watkins, H.; Mayosi, B.M. Arrhythmogenic right ventricular cardiomyopathy type 6 (*arvc6*): Support for the locus assignment, narrowing of the critical region and mutation screening of three candidate genes. *BMC Med. Genet.* **2006**, *7*. [[CrossRef](#)] [[PubMed](#)]

92. Melberg, A.; Oldfors, A.; Blomstrom-Lundqvist, C.; Stalberg, E.; Carlsson, B.; Larrson, E.; Lidell, C.; Eeg-Olofsson, K.E.; Wikstrom, G.; Henriksson, G.; et al. Autosomal dominant myofibrillar myopathy with arrhythmogenic right ventricular cardiomyopathy linked to chromosome 10q. *Ann. Neurol.* **1999**, *46*, 684–692. [[CrossRef](#)]
93. Basso, C.; Fox, P.R.; Meurs, K.M.; Towbin, J.A.; Spier, A.W.; Calabrese, F.; Maron, B.J.; Thiene, G. Arrhythmogenic right ventricular cardiomyopathy causing sudden cardiac death in boxer dogs: A new animal model of human disease. *Circulation* **2004**, *109*, 1180–1185. [[CrossRef](#)] [[PubMed](#)]
94. Meurs, K.M.; Mauceli, E.; Lahmers, S.; Acland, G.M.; White, S.N.; Lindblad-Toh, K. Genome-wide association identifies a deletion in the 3' untranslated region of striatin in a canine model of arrhythmogenic right ventricular cardiomyopathy. *Hum. Genet.* **2010**, *128*, 315–324. [[CrossRef](#)] [[PubMed](#)]
95. Simpson, M.A.; Cook, R.W.; Solanki, P.; Patton, M.A.; Dennis, J.A.; Crosby, A.H. A mutation in nfkB interacting protein 1 causes cardiomyopathy and woolly haircoat syndrome of poll hereford cattle. *Anim. Genet.* **2009**, *40*, 42–46. [[CrossRef](#)] [[PubMed](#)]
96. Zhou, B.; Rao, L.; Peng, Y.; Wang, Y.; Li, Y.; Gao, L.; Chen, Y.; Xue, H.; Song, Y.; Liao, M.; et al. Functional polymorphism of the nfkB1 gene promoter is related to the risk of dilated cardiomyopathy. *BMC Med. Genet.* **2009**, *10*. [[CrossRef](#)] [[PubMed](#)]
97. Fox, P.R.; Maron, B.J.; Basso, C.; Liu, S.-K.; Thiene, G. Spontaneously occurring arrhythmogenic right ventricular cardiomyopathy in the domestic cat. *New Anim. Model Similar Hum. Dis.* **2000**, *102*, 1863–1870.
98. Freel, K.M.; Morrison, L.R.; Thompson, H.; Else, R.W. Arrhythmogenic right ventricular cardiomyopathy as a cause of unexpected cardiac death in two horses. *Vet. Record* **2010**, *166*, 718–722. [[CrossRef](#)] [[PubMed](#)]
99. Raftery, A.G.; Garcia, N.C.; Thompson, H.; Sutton, D.G.M. Arrhythmogenic right ventricular cardiomyopathy secondary to adipose infiltration as a cause of episodic collapse in a horse. *Ir. Vet. J.* **2015**, *68*. [[CrossRef](#)]
100. Cohen, N.; Muntoni, F. Multiple pathogenetic mechanisms in x linked dilated cardiomyopathy. *Heart* **2004**, *90*, 835–841. [[CrossRef](#)] [[PubMed](#)]
101. Chamberlain, R.C.; Smith, E.C.; Campbell, M.J. Novel rod domain duplication in dystrophin resulting in x-linked dilated cardiomyopathy. *Pediatr. Neurol.* **2015**, *53*, 439–441. [[CrossRef](#)] [[PubMed](#)]
102. Nakamura, A. X-linked dilated cardiomyopathy: A cardiospecific phenotype of dystrophinopathy. *Pharmaceuticals* **2015**, *8*, 303–320. [[CrossRef](#)] [[PubMed](#)]
103. D'Arcy, C.; Kanellakis, V.; Forbes, R.; Wilding, B.; McGrath, M.; Howell, K.; Ryan, M.; McLean, C. X-linked recessive distal myopathy with hypertrophic cardiomyopathy caused by a novel mutation in the fhl1 gene. *J. Child. Neurol.* **2015**, *30*, 1211–1217. [[CrossRef](#)] [[PubMed](#)]
104. Bione, S.; D'Adamo, P.; Maestrini, E.; Gedeon, A.K.; Bolhuis, P.A.; Toniolo, D. A novel x-linked gene, g4.5. Is responsible for barth syndrome. *Nat. Genet.* **1996**, *12*, 385–389. [[CrossRef](#)] [[PubMed](#)]
105. Bione, S.; Maestrini, E.; Rivella, S.; Mancini, M.; Regis, S.; Romeo, G.; Toniolo, D. Identification of a novel x-linked gene responsible for emery-dreifuss muscular-dystrophy. *Nat. Genet.* **1994**, *8*, 323–327. [[CrossRef](#)] [[PubMed](#)]
106. Nishino, I.; Fu, J.; Tanji, K.; Yamada, T.; Shimojo, S.; Koori, T.; Mora, M.; Riggs, J.E.; Oh, S.J.; Koga, Y.; et al. Primary lamp-2 deficiency causes x-linked vacuolar cardiomyopathy and myopathy (danon disease). *Nature* **2000**, *406*, 906–910. [[CrossRef](#)] [[PubMed](#)]
107. Ho, M.F.; Chelly, J.; Carter, N.; Danek, A.; Crocker, P.; Monaco, A.P. Isolation of the gene for mcLeod syndrome that encodes a novel membrane-transport protein. *Cell* **1994**, *77*, 869–880. [[CrossRef](#)]
108. Hoffman, E.P.; Brown, R.H.; Kunkel, L.M. Dystrophin—The protein product of the duchenne muscular-dystrophy locus. *Cell* **1987**, *51*, 919–928. [[CrossRef](#)]
109. Zeviani, M.; Taroni, F. Mitochondrial diseases. *Baillieres Clin. Neurol.* **1994**, *3*, 315–334. [[PubMed](#)]
110. Wang, Q.; Liao, Y.; Gong, F.; Mao, H.; Zhang, J. Possible association of hla-drb1 gene with the autoantibody against myocardial mitochondria adp/atp carrier in dilated cardiomyopathy. *J. Huazhong Univ. Sci. Technol. Med. Sci.* **2002**, *22*, 231–232, 245.
111. Terasaki, F.; Tanaka, M.; Kawamura, K.; Kanzaki, Y.; Okabe, M.; Hayashi, T.; Shimomura, H.; Ito, T.; Suwa, M.; Gong, J.S.; et al. A case of cardiomyopathy showing progression from the hypertrophic to the dilated form: Association of mt8348A->G mutation in the mitochondrial trna(lys) gene with severe ultrastructural alterations of mitochondria in cardiomyocytes. *Jpn. Circ. J.* **2001**, *65*, 691–694. [[CrossRef](#)] [[PubMed](#)]
112. Papadopoulou, L.C.; Theophilidis, G.; Thomopoulos, G.N.; Tsiftoglou, A.S. Structural and functional impairment of mitochondria in adriamycin-induced cardiomyopathy in mice: Suppression of cytochrome c oxidase ii gene expression. *Biochem. Pharmacol.* **1999**, *57*, 481–489. [[CrossRef](#)]

113. Sliwa, K.; Hilfiker-Kleiner, D.; Petrie, M.C.; Mebazaa, A.; Pieske, B.; Buchmann, E.; Regitz-Zagrosek, V.; Schaufelberger, M.; Tavazzi, L.; van Veldhuisen, D.J.; et al. Current state of knowledge on aetiology, diagnosis, management, and therapy of peripartum cardiomyopathy: A position statement from the heart failure association of the european society of cardiology working group on peripartum cardiomyopathy. *Eur. J. Heart Fail.* **2010**, *12*, 767–778. [[CrossRef](#)] [[PubMed](#)]
114. Lacuata, A.Q.; Yamada, H.; Hirose, T. Atrial-fibrillation (af) in a cow with postpartum cardiomyopathy (ppcm)—Case-report. *Philipp J. Vet. Med.* **1980**, *19*, 97.
115. Sandusky, G.E.; Cho, D.Y. Congestive cardiomyopathy in a dog associated with pregnancy. *Cornell Vet.* **1984**, *74*, 60–64. [[PubMed](#)]
116. Bollen, I.A.E.; Van Deel, E.D.; Kuster, D.W.D.; Van Der Velden, J. Peripartum cardiomyopathy and dilated cardiomyopathy: Different at heart. *Front. Physiol.* **2015**, *5*. [[CrossRef](#)] [[PubMed](#)]
117. Hilfiker-Kleiner, D.; Struman, I.; Luchtefeld, M.; Forster, O.; Sliwa, K.; Drexler, H. A cathepsin d-cleaved 16kda form of prolactin mediates postpartum cardiomyopathy: Inhibition of prolactin as a novel therapy option. *Circulation* **2006**, *114*, 89.
118. Van Spaendonck-Zwarts, K.Y.; van Tintelen, J.P.; van Veldhuisen, D.J.; van der Werf, R.; Jongbloed, J.D.H.; Paulus, W.J.; Dooijes, D.; van den Berg, M.P. Peripartum cardiomyopathy as a part of familial dilated cardiomyopathy. *Circulation* **2010**, *121*, 2169–2175. [[CrossRef](#)] [[PubMed](#)]
119. Ware, J.S.; Li, J.; Mazaika, E.; Yasso, C.M.; DeSouza, T.; Cappola, T.P.; Tsai, E.J.; Hilfiker-Kleiner, D.; Kamiya, C.A.; Mazarotto, F.; et al. Shared genetic predisposition in peripartum and dilated cardiomyopathies. *New Engl. J. Med.* **2016**, *374*, 233–241. [[CrossRef](#)] [[PubMed](#)]
120. Hershberger, R.E.; Siegfried, J.D. Update 2011: Clinical and genetic issues in familial dilated cardiomyopathy. *J. Am. Coll. Cardiol.* **2011**, *57*, 1641–1649. [[CrossRef](#)] [[PubMed](#)]
121. Amador, F.J.; Kimlicka, L.; Stathopoulos, P.B.; Gasmi-Seabrook, G.M.; MacLennan, D.H.; Van Petegem, F.; Ikura, M. Type 2 ryanodine receptor domain a contains a unique and dynamic alpha-helix that transitions to a beta-strand in a mutant linked with a heritable cardiomyopathy. *J. Mol. Biol.* **2013**, *425*, 4034–4046. [[CrossRef](#)] [[PubMed](#)]
122. GenomicsEngland. The 100,000 Genome Project. Available online: <https://www.genomicsengland.co.uk/the-100000-genomes-project/> (accessed on 28 December 2016).
123. Stedmand, N.L.; Brow, T.P. Cardiomyopathy in broiler chickens congenitally infected with avian leukosis virus subgroup. *J. Vet. Pathol.* **2002**, *39*, 161–164. [[CrossRef](#)] [[PubMed](#)]
124. Simpson, S.; Dunning, M.D.; Brownlie, S.; Patel, J.; Godden, M.; Cobb, M.; Mongan, N.P.; Rutland, C.S. Multiple genetic associations with Irish wolfhound dilated cardiomyopathy. *BioMed Res. Int.* **2016**, *3*. [[CrossRef](#)] [[PubMed](#)]
125. Simpson, S.; Edwards, J.; Emes, R.D.; Cobb, M.A.; Mongan, N.P.; Rutland, C.S. A predictive model for canine dilated cardiomyopathy—a meta-analysis of doberman pinscher data. *PeerJ* **2015**, *3*. [[CrossRef](#)] [[PubMed](#)]
126. HealthInCode. Medical Genetics. Available online: www.healthincode.com (accessed on 28 December 2016).
127. Bienengraeber, M.; Olson, T.M.; Selivanov, V.A.; Kathmann, E.C.; O’Coilain, F.; Gao, F.; Karger, A.B.; Ballew, J.D.; Hodgson, D.M.; Zingman, L.V.; et al. Abcc9 mutations identified in human dilated cardiomyopathy disrupt catalytic katp channel gating. *Nat. Genet.* **2004**, *36*, 382–387. [[CrossRef](#)] [[PubMed](#)]
128. Olson, T.M.; Michels, V.V.; Thibodeau, S.N.; Tai, Y.S.; Keating, M.T. Actin mutations in dilated cardiomyopathy, a heritable form of heart failure. *Science* **1998**, *280*, 750–752. [[CrossRef](#)] [[PubMed](#)]
129. Mogensen, J.; Klausen, I.C.; Pedersen, A.K.; Egeblad, H.; Bross, P.; Kruse, T.A.; Gregersen, N.; Hansen, P.S.; Baandrup, U.; Borlum, A.D. Alpha-cardiac actin is a novel disease gene in familial hypertrophic cardiomyopathy. *J. Clin. Investig.* **1999**, *103*, R39–R43. [[CrossRef](#)] [[PubMed](#)]
130. Chiu, C.; Bagnall, R.D.; Ingles, J.; Yeates, L.; Kennerson, M.; Donald, J.A.; Jormakka, M.; Lind, J.M.; Semsarian, C. Mutations in alpha-actinin-2 cause hypertrophic cardiomyopathy a genome-wide analysis. *J. Am. Coll. Cardiol.* **2010**, *55*, 1127–1135. [[CrossRef](#)] [[PubMed](#)]
131. Mohapatra, B.; Jimenez, S.; Lin, J.H.; Bowles, K.R.; Coveler, K.J.; Marx, J.G.; Chrisco, M.A.; Murphy, R.T.; Lurie, P.R.; Schwartz, R.J.; et al. Mutations in the muscle lim protein and alpha-actinin-2 genes in dilated cardiomyopathy and endocardial fibroelastosis. *Mol. Genet. Metab.* **2003**, *80*, 207–215. [[CrossRef](#)]
132. Moulik, M.; Vatta, M.; Witt, S.H.; Arola, A.M.; Murphy, R.T.; McKenna, W.J.; Boriek, A.M.; Oka, K.; Labeit, S.; Bowles, N.E.; et al. Ankrd1, the gene encoding cardiac ankyrin repeat protein, is a novel dilated cardiomyopathy gene. *J. Am. Coll. Cardiol.* **2009**, *54*, 325–333. [[CrossRef](#)] [[PubMed](#)]

133. Arimura, T.; Bos, J.M.; Sato, A.; Kubo, T.; Okamoto, H.; Nishi, H.; Harada, H.; Koga, Y.; Moulik, M.; Doi, Y.L.; et al. Cardiac ankyrin repeat protein gene (ankrd1) mutations in hypertrophic cardiomyopathy. *Hum. Mutat.* **2011**, *32*, 1481–1491. [[CrossRef](#)] [[PubMed](#)]
134. Norton, N.; Li, D.X.; Rieder, M.J.; Siegfried, J.D.; Rampersaud, E.; Zuchner, S.; Mangos, S.; Gonzalez-Quintana, J.; Wang, L.B.; McGee, S.; et al. Genome-wide studies of copy number variation and exome sequencing identify rare variants in bag3 as a cause of dilated cardiomyopathy. *Am. J. Hum. Genet.* **2011**, *88*, 273–282. [[CrossRef](#)] [[PubMed](#)]
135. Villard, E.; Perret, C.; Gary, F.; Proust, C.; Dilanian, G.; Isnard, R.; Komajda, M.; Charron, P.; Cambien, F. A genome-wide association study identifies two loci associated with heart failure due to dilated cardiomyopathy. *Eur. Heart J.* **2011**, *32*, 1065–1076. [[CrossRef](#)] [[PubMed](#)]
136. Arimura, T.; Ishikawa, T.; Nunoda, S.; Kawai, S.; Kimura, A. Dilated cardiomyopathy-associated bag3 mutations impair z-disc assembly and enhance sensitivity to apoptosis in cardiomyocytes. *Hum. Mutat.* **2011**, *32*, 1481–1491. [[CrossRef](#)] [[PubMed](#)]
137. Chiu, C.; Tebo, M.; Ingles, J.; Yeates, L.; Arthur, J.W.; Lind, J.M.; Semsarian, C. Genetic screening of calcium regulation genes in familial hypertrophic cardiomyopathy. *J. Mol. Cell. Cardiol.* **2007**, *43*, 337–343. [[CrossRef](#)] [[PubMed](#)]
138. Catteruccia, M.; Sanna, T.; Santorelli, F.M.; Tessa, A.; Di Giacomo, R.; Sauchelli, D.; Verbo, A.; Lo Monaco, M.; Servidei, S. Rippling muscle disease and cardiomyopathy associated with a mutation in the cav3 gene. *Neuromuscul. Disord.* **2009**, *19*, 779–783. [[CrossRef](#)] [[PubMed](#)]
139. Zhang, L.; Hu, A.H.; Yuan, H.X.; Cui, L.; Miao, G.B.; Yang, X.C.; Wang, L.F.; Liu, J.C.; Liu, X.L.; Wang, S.Y.; et al. A missense mutation in the chrm2 gene is associated with familial dilated cardiomyopathy. *Circ. Res.* **2008**, *102*, 1426–1432. [[CrossRef](#)] [[PubMed](#)]
140. Inagaki, N.; Hayashi, T.; Arimura, T.; Koga, Y.; Takahashi, M.; Shibata, H.; Teraoka, K.; Chikamori, T.; Yamashina, A.; Kimura, A. Alpha b-crystallin mutation in dilated cardiomyopathy. *Biochem. Biophys. Res. Commun.* **2006**, *342*, 379–386. [[PubMed](#)]
141. Bos, J.M.; Poley, R.N.; Ny, M.; Tester, D.J.; Xu, X.L.; Vatta, M.; Towbin, J.A.; Gersh, B.J.; Ommen, S.R.; Ackerman, M.J. Genotype-phenotype relationships involving hypertrophic cardiomyopathy-associated mutations in titin, muscle lim protein, and telethonin. *Mol. Genet. Metab.* **2006**, *88*, 78–85. [[CrossRef](#)] [[PubMed](#)]
142. Erdmann, J.; Hassfeld, S.; Kallisch, H.; Fleck, E.; Regitz-Zagrose, V. Genetic variants in the promoter (g983g>t) and coding region (a92t) of the human cardiotrophin-1 gene (ctf1) in patients with dilated cardiomyopathy. *Hum. Mutat.* **2000**, *16*, 448. [[CrossRef](#)]
143. van Hengel, J.; Calore, M.; Bauce, B.; Dazzo, E.; Mazzotti, E.; De Bortoli, M.; Lorenzon, A.; Li Mura, I.E.A.; Boffagna, G.; Rigato, I.; et al. Mutations in the area composita protein t-catenin are associated with arrhythmogenic right ventricular cardiomyopathy. *Eur. Heart J.* **2013**, *34*, 201–210. [[CrossRef](#)] [[PubMed](#)]
144. Schaper, J.; Froede, R.; Stheun, B.; Buck, A.; Hashizume, H.; Speiser, B.; Friedl, A.; Bleese, N. Impairment of the myocardial ultrastructure and changes of the cytoskeleton in dilated cardiomyopathy. *Circulation* **1991**, *83*, 504–514. [[CrossRef](#)] [[PubMed](#)]
145. Li, D.X.; Tapscoft, T.; Gonzalez, O.; Burch, P.E.; Quinones, M.A.; Zoghbi, W.A.; Hill, R.; Bachinski, L.L.; Mann, D.L.; Roberts, R. Desmin mutation responsible for idiopathic dilated cardiomyopathy. *Circulation* **1999**, *100*, 461–464. [[CrossRef](#)] [[PubMed](#)]
146. Lorenzon, A.; Boffagna, G.; Bauce, B.; De Bortoli, M.; Mura, I.E.A.L.; Calore, M.; Dazzo, E.; Basso, C.; Nava, A.; Thiene, G.; et al. Desmin mutations and arrhythmogenic right ventricular cardiomyopathy. *Am. J. Cardiol.* **2013**, *111*, 400–405. [[CrossRef](#)] [[PubMed](#)]
147. Klauke, B.; Kossmann, S.; Gaertner, A.; Brand, K.; Stork, I.; Brodehl, A.; Dieding, M.; Walhorn, V.; Anselmetti, D.; Gerdes, D.; et al. De novo desmin-mutation n116s is associated with arrhythmogenic right ventricular cardiomyopathy. *Hum. Mol. Genet.* **2010**, *19*, 4595–4607. [[CrossRef](#)] [[PubMed](#)]
148. Muntoni, F.; Cau, M.; Ganau, A.; Congiu, R.; Arvedi, G.; Mateddu, A.; Marrosu, M.G.; Cianchetti, C.; Realdi, G.; Cao, A.; et al. Brief report—Deletion of the dystrophin muscle-promoter region associated with x-linked dilated cardiomyopathy. *N. Engl. J. Med.* **1993**, *329*, 921–925. [[CrossRef](#)] [[PubMed](#)]
149. OrtizLopez, R.; Li, H.; Su, J.; Goytia, V.; Towbin, J.A. Evidence for a dystrophin missense mutation as a cause of x-linked dilated cardiomyopathy. *Circulation* **1997**, *95*, 2434–2440. [[CrossRef](#)]

150. Davey, K.M.; Parboosingh, J.S.; McLeod, D.R.; Chan, A.; Casey, R.; Ferreira, P.; Snyder, F.F.; Bridge, P.J.; Bernier, F.P. Mutation of dnajc19, a human homologue of yeast inner mitochondrial membrane co-chaperones, causes dcma syndrome, a novel autosomal recessive barth syndrome-like condition. *J. Med. Genet.* **2006**, *43*, 385–393. [[CrossRef](#)] [[PubMed](#)]
151. Lefeber, D.J.; de Brouwer, A.P.M.; Morava, E.; Riemersma, M.; Schuurs-Hoeijmakers, J.H.M.; Absmanner, B.; Verrijp, K.; van den Akker, W.M.R.; Huijben, K.; Steenbergen, G.; et al. Autosomal recessive dilated cardiomyopathy due to dolk mutations results from abnormal dystroglycan o-mannosylation. *PLoS Genet.* **2011**, *7*. [[CrossRef](#)] [[PubMed](#)]
152. Beffagna, G.; De Bortoli, M.; Nava, A.; Salamon, M.; Lorenzon, A.; Zaccolo, M.; Mancuso, L.; Sigalotti, L.; Bauce, B.; Occhi, G.; et al. Missense mutations in desmocollin-2 n-terminus, associated with arrhythmogenic right ventricular cardiomyopathy, affect intracellular localization of desmocollin-2 in vitro. *BMC Med. Genet.* **2007**, *8*. [[CrossRef](#)] [[PubMed](#)]
153. Posch, M.G.; Posch, M.J.; Geier, C.; Erdmann, B.; Mueller, W.; Richter, A.; Ruppert, V.; Pankuweit, S.; Maisch, B.; Perrot, A.; et al. A missense variant in desmoglein-2 predisposes to dilated cardiomyopathy. *Mol. Genet. Metab.* **2008**, *95*, 74–80. [[CrossRef](#)] [[PubMed](#)]
154. Pilichou, K.; Nava, A.; Basso, C.; Beffagna, G.; Bauce, B.; Lorenzon, A.; Frigo, G.; Vettori, A.; Valente, M.; Towbin, J.; et al. Mutations in desmoglein-2 gene are associated with arrhythmogenic right ventricular cardiomyopathy. *Circulation* **2006**, *113*, 1171–1179. [[CrossRef](#)] [[PubMed](#)]
155. Awad, M.M.; Dalal, D.; Cho, E.; Amat-Alarcon, N.; James, C.; Tichnell, C.; Tucker, A.; Russell, S.D.; Bluemke, D.A.; Dietz, H.C.; et al. Dsg2 mutations contribute to arrhythmogenic right ventricular dysplasia/cardiomyopathy. *Am. J. Hum. Genet.* **2006**, *79*, 136–142. [[CrossRef](#)] [[PubMed](#)]
156. Norgett, E.E.; Hatsell, S.J.; Carvajal-Huerta, L.; Cabezas, J.C.R.; Common, J.; Purkis, P.E.; Whittock, N.; Leigh, I.M.; Stevens, H.P.; Kelsell, D.P. Recessive mutation in desmoplakin disrupts desmoplakin-intermediate filament interactions and causes dilated cardiomyopathy, woolly hair and keratoderma. *Hum. Mol. Genet.* **2000**, *9*, 2761–2766. [[CrossRef](#)] [[PubMed](#)]
157. Rampazzo, A.; Nava, A.; Malacrida, S.; Beffagna, G.; Bauce, B.; Rossi, V.; Zimbello, R.; Simionati, B.; Basso, C.; Thiene, G.; et al. Mutation in human desmoplakin domain binding to plakoglobin causes a dominant form of arrhythmogenic right ventricular cardiomyopathy. *Am. J. Hum. Genet.* **2002**, *71*, 1200–1206. [[CrossRef](#)] [[PubMed](#)]
158. Zhang, M.Q.; Chen, J.; Si, D.Y.; Zheng, Y.; Jiao, H.X.; Feng, Z.H.; Hu, Z.M.; Duan, R.H. Whole exome sequencing identifies a novel emd mutation in a chinese family with dilated cardiomyopathy. *BMC Med. Genet.* **2014**, *15*. [[CrossRef](#)] [[PubMed](#)]
159. Schonberger, J.; Wang, L.; Shin, T.J.; Kim, S.D.; Depreux, F.F.S.; Zhu, H.; Zon, L.; Pizard, A.; Kim, J.B.; MacRae, C.A.; et al. Mutation in the transcriptional coactivator eya4 causes dilated cardiomyopathy and sensorineural hearing loss. *Nat. Genet.* **2005**, *37*, 418–422. [[CrossRef](#)] [[PubMed](#)]
160. Al-Yacoub, N.; Shaheen, R.; Awad, S.M.; Kunhi, M.; Dzimiri, N.; Nguyen, H.C.; Xiong, Y.; Al-Buraiki, J.; Al-Habeeb, W.; Alkuraya, F.S.; et al. Fbxo32, encoding a member of the scf complex, is mutated in dilated cardiomyopathy. *Genome Biol.* **2016**, *17*. [[CrossRef](#)] [[PubMed](#)]
161. Arimura, T.; Hayashi, T.; Matsumoto, Y.; Shibata, H.; Hiroi, S.; Nakamura, T.; Inagaki, N.; Hinohara, K.; Takahashi, M.; Manatsu, S.I.; et al. Structural analysis of four and half lim protein-2 in dilated cardiomyopathy. *Biochem. Biophys. Res. Commun.* **2007**, *357*, 162–167. [[CrossRef](#)] [[PubMed](#)]
162. Murakami, T.; Hayashi, Y.K.; Noguchi, S.; Ogawa, M.; Nonaka, I.; Tanabe, Y.; Ogino, M.; Takada, F.; Eriguchi, M.; Kotooka, N.; et al. Fukutin gene mutations cause dilated cardiomyopathy with minimal muscle weakness. *Ann. Neurol.* **2006**, *60*, 597–602. [[CrossRef](#)] [[PubMed](#)]
163. Muller, T.; Krasnianski, M.; Witthaut, R.; Deschauer, M.; Zierz, S. Dilated cardiomyopathy may be an early sign of the c826a fukutin-related protein mutation. *Neuromusc. Disord.* **2005**, *15*, 372–376. [[CrossRef](#)] [[PubMed](#)]
164. Minoretti, P.; Arra, M.; Emanuele, E.; Olivieri, V.; Aldeghi, A.; Politi, P.; Martinelli, V.; Pesenti, S.; Falcone, C. A w148r mutation in the human foxd4 gene segregating with dilated cardiomyopathy, obsessive-compulsive disorder, and suicidality. *Int. J. Mol. Med.* **2007**, *19*, 369–372. [[CrossRef](#)] [[PubMed](#)]
165. Theis, J.L.; Sharpe, K.M.; Matsumoto, M.E.; Chai, H.S.; Nair, A.A.; Theis, J.D.; de Andrade, M.; Wieben, E.D.; Michels, V.V.; Olson, T.M. Homozygosity mapping and exome sequencing reveal gatad1 mutation in autosomal recessive dilated cardiomyopathy. *Circ. Cardiovasc. Genet.* **2011**, *4*, 585–644. [[CrossRef](#)] [[PubMed](#)]

166. Zhou, Y.M.; Dai, X.Y.; Qiu, X.B.; Yuan, F.; Li, R.G.; Xu, Y.J.; Qu, X.K.; Huang, R.T.; Xue, S.; Yang, Y.Q. Hand1 loss-of-function mutation associated with familial dilated cardiomyopathy. *Clin. Chem. Lab. Med.* **2016**, *54*, 1161–1167. [[CrossRef](#)] [[PubMed](#)]
167. Pankuweit, S.; Ruppert, V.; Jonsdottir, P.; Muller, H.H.; Meyer, T.; Heart, G.C.N. The hla class ii allele dqb1*0309 is associated with dilated cardiomyopathy. *Gene* **2013**, *531*, 180–183. [[CrossRef](#)] [[PubMed](#)]
168. Stark, K.; Esslinger, U.B.; Reinhard, W.; Petrov, G.; Winkler, T.; Komajda, M.; Isnard, R.; Charron, P.; Villard, E.; Cambien, F.; et al. Genetic association study identifies hspb7 as a risk gene for idiopathic dilated cardiomyopathy. *PLoS Genet.* **2010**, *6*. [[CrossRef](#)] [[PubMed](#)]
169. Knoll, R.; Postel, R.; Wang, J.; Kratzner, R.; Hennecke, G.; Vacaru, A.M.; Vakeel, P.; Schubert, C.; Murthy, K.; Rana, B.K.; et al. Laminin-alpha4 and integrin-linked kinase mutations cause human cardiomyopathy via simultaneous defects in cardiomyocytes and endothelial cells. *Circulation* **2007**, *116*, 515–525. [[CrossRef](#)] [[PubMed](#)]
170. Landstrom, A.P.; Weisleder, N.; Bataalden, K.B.; Bos, J.M.; Tester, D.J.; Ommen, S.R.; Wehrens, X.H.; Claycomb, W.C.; Ko, J.K.; Hwang, M.; et al. Mutations in jph2-encoded junctophilin-2 associated with hypertrophic cardiomyopathy in humans. *J. Mol. Cell. Cardiol.* **2007**, *42*, 1026–1035. [[CrossRef](#)] [[PubMed](#)]
171. McKoy, G.; Protonotarios, N.; Crosby, A.; Tsatsopoulou, A.; Anastasakis, A.; Coonar, A.; Norman, M.; Baboonian, C.; Jeffery, S.; McKenna, W.J. Identification of a deletion in plakoglobin in arrhythmogenic right ventricular cardiomyopathy with palmoplantar keratoderma and woolly hair (naxos disease). *Lancet* **2000**, *355*, 2119–2124. [[CrossRef](#)]
172. Carboni, N.; Marrosu, G.; Porcu, M.; Mateddu, A.; Solla, E.; Cocco, E.; Maioli, M.A.; Oppo, V.; Piras, R.; Marrosu, M.G. Dilated cardiomyopathy with conduction defects in a patient with partial merosin deficiency due to mutations in the laminin-alpha 2-chain gene: A chance association or a novel phenotype? *Muscle Nerve* **2011**, *44*, 826–828. [[CrossRef](#)] [[PubMed](#)]
173. Maron, B.J.; Roberts, W.C.; Arad, M.; Haas, T.S.; Spirito, P.; Wright, G.B.; Almquist, A.K.; Baffa, J.M.; Saul, J.P.; Ho, C.Y.; et al. Clinical outcome and phenotypic expression in lamp2 cardiomyopathy. *JAMA* **2009**, *301*, 1253–1259. [[CrossRef](#)] [[PubMed](#)]
174. Arimura, T.; Hayashi, T.; Terada, H.; Lee, S.Y.; Zhou, Q.; Takahashi, M.; Ueda, K.; Nouchi, T.; Hohda, S.; Shibutani, M.; et al. A cypher/zasp mutation associated with dilated cardiomyopathy alters the binding affinity to protein kinase c. *J. Biol. Chem.* **2004**, *279*, 6746–6752. [[CrossRef](#)] [[PubMed](#)]
175. Garcia-Pavia, P.; Vazquez, M.E.; Segovia, J.; Salas, C.; Avellana, P.; Gomez-Bueno, M.; Vilches, C.; Gallardo, M.E.; Garesse, R.; Molano, J.; et al. Genetic basis of end-stage hypertrophic cardiomyopathy. *Eur. J. Heart Fail.* **2011**, *13*, 1193–1201. [[CrossRef](#)] [[PubMed](#)]
176. Fatkin, D.; MacRae, C.; Sasaki, T.; Wolff, M.R.; Porcu, M.; Frenneaux, M.; Atherton, J.; Vidaillet, H.J.; Spudich, S.; De Girolami, U.; et al. Missense mutations in the rod domain of the lamin a/c gene as causes of dilated cardiomyopathy and conduction-system disease. *N. Engl. J. Med.* **1999**, *341*, 1715–1724. [[CrossRef](#)] [[PubMed](#)]
177. Quarta, G.; Syrris, P.; Ashworth, M.; Jenkins, S.; Alapi, K.Z.; Morgan, J.; Muir, A.; Pantazis, A.; McKenna, W.J.; Elliott, P.M. Mutations in the lamin a/c gene mimic arrhythmogenic right ventricular cardiomyopathy. *Eur. Heart J.* **2012**, *33*, 1128–1149. [[CrossRef](#)] [[PubMed](#)]
178. Qu, X.K.; Yuan, F.; Li, R.G.; Xu, L.; Jing, W.F.; Liu, H.; Xu, Y.J.; Zhang, M.; Liu, X.; Fang, W.Y.; et al. Prevalence and spectrum of lrcc10 mutations associated with idiopathic dilated cardiomyopathy. *Mol. Med. Rep.* **2015**, *12*, 3718–3724. [[PubMed](#)]
179. Rodriguez, G.; Ueyama, T.; Ogata, T.; Czernuszewicz, G.; Tan, Y.L.; Dorn, G.W.; Bogaev, R.; Amano, K.; Oh, H.; Matsubara, H.; et al. Molecular genetic and functional characterization implicate muscle-restricted coiled-coil gene (murc) as a causal gene for familial dilated cardiomyopathy. *Circ. Cardiovasc. Genet.* **2011**, *4*, 349–357. [[CrossRef](#)] [[PubMed](#)]
180. Daehmlow, S.; Erdmann, J.; Knueppel, T.; Gille, C.; Froemmel, C.; Hummel, M.; Hetzer, R.; Regitz-Zagrosek, V. Novel mutations in sarcomeric protein genes in dilated cardiomyopathy. *Biochem. Biophys. Res. Commun.* **2002**, *298*, 116–120. [[CrossRef](#)]
181. Carrier, L.; Bonne, G.; Bahrend, E.; Yu, B.; Richard, P.; Niel, F.; Hainque, B.; Cruaud, C.; Gary, F.; Labeit, S.; et al. Organization and sequence of human cardiac myosin binding protein c gene (mybpc3) and identification of mutations predicted to produce truncated proteins in familial hypertrophic cardiomyopathy. *Circ. Res.* **1997**, *80*, 427–434. [[PubMed](#)]

182. Bonne, G.; Carrier, L.; Bercovici, J.; Cruaud, C.; Richard, P.; Hainque, B.; Gautel, M.; Labeit, S.; James, M.; Beckmann, J.; et al. Cardiac myosin binding protein-c gene splice acceptor site mutation is associated with familial hypertrophic cardiomyopathy. *Nat. Genet.* **1995**, *11*, 438–440. [[CrossRef](#)] [[PubMed](#)]
183. Hershberger, R.E.; Norton, N.; Morales, A.; Li, D.X.; Siegfried, J.D.; Gonzalez-Quintana, J. Coding sequence rare variants identified in mybpc3, myh6, tpm1, tnnc1, and tnni3 from 312 patients with familial or idiopathic dilated cardiomyopathy. *Circ. Cardiovasc. Genet.* **2010**, *3*, 155–161. [[CrossRef](#)] [[PubMed](#)]
184. Niimura, H.; Patton, K.K.; McKenna, W.J.; Soultis, J.; Maron, B.J.; Seidman, J.G.; Seidman, C.E. Sarcomere protein gene mutations in hypertrophic cardiomyopathy of the elderly. *Circulation* **2002**, *105*, 446–451. [[CrossRef](#)] [[PubMed](#)]
185. Fananapazir, L.; Dalakas, M.C.; Cyran, F.; Cohn, G.; Epstein, N.D. Missense mutations in the beta-myosin heavy-chain gene cause central core disease in hypertrophic cardiomyopathy. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 3993–3997. [[CrossRef](#)] [[PubMed](#)]
186. Karkkainen, S.; Helio, T.; Jaaskelainen, P.; Miettinen, R.; Tuomainen, P.; Ylitalo, K.; Kaartinen, M.; Reissell, E.; Toivonen, L.; Nieminen, M.S.; et al. Two novel mutations in the beta-myosin heavy chain gene associated with dilated cardiomyopathy. *Eur. J. Heart Fail.* **2004**, *6*, 861–868. [[CrossRef](#)] [[PubMed](#)]
187. Flavigny, J.; Richard, P.; Isnard, R.; Carrier, L.; Charron, P.; Bonne, G.; Forissier, J.F.; Desnos, M.; Dubourg, O.; Komajda, M.; et al. Identification of two novel mutations in the ventricular regulatory myosin light chain gene (myl2) associated with familial and classical forms of hypertrophic cardiomyopathy. *J. Mol. Med.* **1998**, *76*, 208–214. [[CrossRef](#)] [[PubMed](#)]
188. Ingles, J.; Doolan, A.; Chiu, C.; Seidman, J.; Seidman, C.; Semsarian, C. Compound and double mutations in patients with hypertrophic cardiomyopathy: Implications for genetic testing and counselling. *J. Med. Genet.* **2005**, *42*. [[CrossRef](#)] [[PubMed](#)]
189. Osio, A.; Tan, L.; Chen, S.N.; Lombardi, R.; Nagueh, S.F.; Shete, S.; Roberts, R.; Willerson, J.T.; Marian, A.J. Myozenin 2 is a novel gene for human hypertrophic cardiomyopathy. *Circ. Res.* **2007**, *100*, 766–768. [[CrossRef](#)] [[PubMed](#)]
190. Duboscq-Bidot, L.; Xu, P.; Charron, P.; Neyroud, N.; Dilanian, G.; Millaire, A.; Bors, V.; Komajda, M.; Villard, E. Mutations in the z-band protein myopalladin gene and idiopathic dilated cardiomyopathy. *Cardiovasc. Res.* **2008**, *77*, 118–125. [[CrossRef](#)] [[PubMed](#)]
191. Purevjav, E.; Varela, J.; Morgado, M.; Kearney, D.L.; Li, H.; Taylor, M.D.; Arimura, T.; Moncman, C.L.; McKenna, W.; Murphy, R.T.; et al. Nebulette mutations are associated with dilated cardiomyopathy and endocardial fibroelastosis. *J. Am. Coll. Cardiol.* **2010**, *56*, 1493–1502. [[CrossRef](#)] [[PubMed](#)]
192. Hassel, D.; Dahme, T.; Erdmann, J.; Meder, B.; Hugel, A.; Stoll, M.; Just, S.; Hess, A.; Ehlermann, P.; Weichenhan, D.; et al. Nexilin mutations destabilize cardiac z-disks and lead to dilated cardiomyopathy. *Nat. Med.* **2009**, *15*. [[CrossRef](#)] [[PubMed](#)]
193. Matsa, L.S.; Rangaraju, A.; Vengaldas, V.; Latifi, M.; Jahromi, H.M.; Ananthapur, V.; Nallari, P. Haplotypes of nos3 gene polymorphisms in dilated cardiomyopathy. *PLoS ONE* **2013**, *8*. [[CrossRef](#)] [[PubMed](#)]
194. Cruz, F.M.; Sanz-Rosa, D.; Roche-Molina, M.; Garcia-Prieto, J.; Garcia-Ruiz, J.M.; Pizarro, G.; Jimenez-Borreguero, L.J.; Torres, M.; Bernad, A.; Ruiz-Cabello, J.; et al. Exercise triggers arvc phenotype in mice expressing a disease-causing mutated version of human plakophilin-2. *J. Am. Coll. Cardiol.* **2015**, *65*, 1438–1450. [[PubMed](#)]
195. Muhammad, E.; Levitas, A.; Singh, S.R.; Braiman, A.; Ofir, R.; Etzion, S.; Sheffield, V.C.; Etzion, Y.; Carrier, L.; Parvari, R. Plekhm2 mutation leads to abnormal localization of lysosomes, impaired autophagy flux and associates with recessive dilated cardiomyopathy and left ventricular noncompaction. *Hum. Mol. Genet.* **2015**, *24*, 7227–7240. [[CrossRef](#)] [[PubMed](#)]
196. Haghghi, K.; Kolokathis, F.; Pater, L.; Lynch, R.A.; Asahi, M.; Gramolini, A.O.; Fan, G.C.; Tsiapras, D.; Hahn, H.S.; Adamopoulos, S.; et al. Human phospholamban null results in lethal dilated cardiomyopathy revealing a critical difference between mouse and human. *J. Clin. Investig.* **2003**, *111*, 869–876. [[CrossRef](#)] [[PubMed](#)]
197. Minamisawa, S.; Sato, Y.; Tatsuguchi, Y.; Fujino, T.; Imamura, S.; Uetsuka, Y.; Nakazawa, M.; Matsuoka, R. Mutation of the phospholamban promoter associated with hypertrophic cardiomyopathy. *Biochem. Biophys. Res. Commun.* **2003**, *304*, 1–4. [[CrossRef](#)]

198. Van der Zwaag, P.A.; van Rijsingen, I.A.W.; Asimaki, A.; Jongbloed, J.D.H.; van Veldhuisen, D.J.; Wiesfeld, A.C.P.; Cox, M.G.P.J.; van Lochem, L.T.; de Boer, R.A.; Hofstra, R.M.W.; et al. Phospholamban r14del mutation in patients diagnosed with dilated cardiomyopathy or arrhythmogenic right ventricular cardiomyopathy: Evidence supporting the concept of arrhythmogenic cardiomyopathy. *Eur. J. Heart Fail.* **2012**, *14*, 1199–1207. [[CrossRef](#)] [[PubMed](#)]
199. Verhoeven, W.M.; Egger, J.I.; Kremer, B.P.; de Pont, B.J.; Marcelis, C.L. Recurrent major depression, ataxia, and cardiomyopathy: Association with a novel polg mutation? *Neuropsychiatr. Dis. Treat.* **2011**, *7*, 293–296. [[CrossRef](#)] [[PubMed](#)]
200. Arndt, A.K.; Schafer, S.; Drenckhahn, J.D.; Sabeh, M.K.; Plovie, E.R.; Caliebe, A.; Klopocki, E.; Musso, G.; Werdich, A.A.; Kalwa, H.; et al. Fine mapping of the 1p36 deletion syndrome identifies mutation of prdm16 as a cause of cardiomyopathy. *Am. J. Hum. Genet.* **2013**, *93*, 67–77. [[CrossRef](#)] [[PubMed](#)]
201. Zhao, Y.; Cao, H.; Song, Y.D.; Feng, Y.; Ding, X.X.; Pang, M.J.; Zhang, Y.M.; Zhang, H.; Ding, J.H.; Xia, X.S. Identification of novel mutations including a double mutation in patients with inherited cardiomyopathy by a targeted sequencing approach using the ion torrent pgm system. *Int. J. Mol. Med.* **2016**, *37*, 1511–1520. [[CrossRef](#)] [[PubMed](#)]
202. Li, D.X.; Parks, S.B.; Kushner, J.D.; Nauman, D.; Burgess, D.; Ludwigsen, S.; Partain, J.; Nixon, R.R.; Allen, C.N.; Irwin, R.P.; et al. Mutations of presenilin genes in dilated cardiomyopathy and heart failure. *Am. J. Hum. Genet.* **2006**, *79*, 1030–1039. [[CrossRef](#)] [[PubMed](#)]
203. Brauch, K.M.; Karst, M.L.; Herron, K.J.; de Andrade, M.; Pellikka, P.A.; Rodeheffer, R.J.; Michels, V.V.; Olson, T.M. Mutations in ribonucleic acid binding protein gene cause familial dilated cardiomyopathy. *J. Am. Coll. Cardiol.* **2009**, *54*, 930–941. [[CrossRef](#)] [[PubMed](#)]
204. Hussain, S.; Haroon, J.; Ejaz, S.; Javed, Q. Variants of resistin gene and the risk of idiopathic dilated cardiomyopathy in pakistan. *Meta Gene* **2016**, *9*, 37–41. [[CrossRef](#)] [[PubMed](#)]
205. Gupta, A.; Colmenero, I.; Ragge, N.K.; Blakely, E.L.; He, L.; McFarland, R.; Taylor, R.W.; Vogt, J.; Milford, D.V. Compound heterozygous rnmnd1 gene variants associated with chronic kidney disease, dilated cardiomyopathy and neurological involvement: A case report. *BMC Res. Notes* **2016**, *9*, 325. [[CrossRef](#)] [[PubMed](#)]
206. Long, P.A.; Zimmermann, M.T.; Kim, M.; Evans, J.M.; Xu, X.; Olson, T.M. De novo rragc mutation activates mtorc1 signaling in syndromic fetal dilated cardiomyopathy. *Hum. Genet.* **2016**, *135*, 909–917. [[CrossRef](#)] [[PubMed](#)]
207. Bhuiyan, Z.A.; van den Berg, M.P.; van Tintelen, J.P.; Bink-Boelkens, M.T.; Wiesfeld, A.C.; Alders, M.; Postma, A.V.; van Langen, I.; Mannens, M.M.; Wilde, A.A. Expanding spectrum of human ryr2-related disease: New electrocardiographic, structural, and genetic features. *Circulation* **2007**, *116*, 1569–1576. [[CrossRef](#)] [[PubMed](#)]
208. Tiso, N.; Stephan, D.A.; Nava, A.; Bagattin, A.; Devaney, J.M.; Stanchi, F.; Larderet, G.; Brahmabhatt, B.; Brown, K.; Bauce, B.; et al. Identification of mutations in the cardiac ryanodine receptor gene in families affected with arrhythmogenic right ventricular cardiomyopathy type 2 (arvd2). *Hum. Mol. Genet.* **2001**, *10*, 189–194. [[CrossRef](#)] [[PubMed](#)]
209. Roux-Buisson, N.; Gandjbakhch, E.; Donal, E.; Probst, V.; Deharo, J.C.; Chevalier, P.; Klug, D.; Mansencal, N.; Delacretaz, E.; Cosnay, P.; et al. Prevalence and significance of rare ryr2 variants in arrhythmogenic right ventricular cardiomyopathy/dysplasia: Results of a systematic screening. *Heart Rhythm* **2014**, *11*, 1999–2009. [[CrossRef](#)] [[PubMed](#)]
210. McNair, W.P.; Ku, L.; Taylor, M.R.; Fain, P.R.; Dao, D.; Wolfel, E.; Mestroni, L.; Familial Cardiomyopathy Registry Research Group. Scn5a mutation associated with dilated cardiomyopathy, conduction disorder, and arrhythmia. *Circulation* **2004**, *110*, 2163–2167. [[CrossRef](#)] [[PubMed](#)]
211. Levitas, A.; Muhammad, E.; Harel, G.; Saada, A.; Caspi, V.C.; Manor, E.; Beck, J.C.; Sheffield, V.; Parvari, R. Familial neonatal isolated cardiomyopathy caused by a mutation in the flavoprotein subunit of succinate dehydrogenase. *Eur. J. Hum. Genet.* **2010**, *18*, 1160–1165. [[CrossRef](#)] [[PubMed](#)]
212. Tsubata, S.; Bowles, K.R.; Vatta, M.; Zintz, C.; Titus, J.; Muhonen, L.; Bowles, N.E.; Towbin, J.A. Mutations in the human delta-sarcoglycan gene in familial and sporadic dilated cardiomyopathy. *J. Clin. Investig.* **2000**, *106*, 655–662. [[CrossRef](#)] [[PubMed](#)]
213. Zhang, Q.P.; Bethmann, C.; Worth, N.F.; Davies, J.D.; Wasner, C.; Feuer, A.; Ragnauth, C.D.; Yi, Q.J.; Mellad, J.A.; Warren, D.T.; et al. Nesprin-1 and -2 are involved in the pathogenesis of emery-dreifuss muscular dystrophy and are critical for nuclear envelope integrity. *Hum. Mol. Genet.* **2007**, *16*, 2816–2833. [[CrossRef](#)] [[PubMed](#)]

214. Kirk, E.P.; Sunde, M.; Costa, M.W.; Rankin, S.A.; Wolstein, O.; Castro, M.L.; Butler, T.L.; Hyun, C.; Guo, G.; Otway, R.; et al. Mutations in cardiac t-box factor gene *tbx20* are associated with diverse cardiac pathologies, including defects of septation and valvulogenesis and cardiomyopathy. *Am. J. Hum. Genet.* **2007**, *81*, 280–291. [[CrossRef](#)] [[PubMed](#)]
215. Zhou, W.; Zhao, L.; Jiang, J.Q.; Jiang, W.F.; Yang, Y.Q.; Qiu, X.B. A novel *tbx5* loss-of-function mutation associated with sporadic dilated cardiomyopathy. *Int. J. Mol. Med.* **2015**, *36*, 282–288. [[CrossRef](#)] [[PubMed](#)]
216. Hayashi, T.; Arimura, T.; Itoh-Satoh, M.; Ueda, K.; Hohda, S.; Inagaki, N.; Takahashi, M.; Hori, H.; Yasunami, M.; Nishi, H.; et al. *Tcap* gene mutations in hypertrophic cardiomyopathy and dilated cardiomyopathy. *J. Am. Coll. Cardiol.* **2004**, *44*, 2192–2201. [[CrossRef](#)] [[PubMed](#)]
217. Beggagna, G.; Occhi, G.; Nava, A.; Vitiello, L.; Ditadi, A.; Basso, C.; Bauce, B.; Carraro, G.; Thiene, G.; Towbin, J.A.; et al. Regulatory mutations in transforming growth factor-beta3 gene cause arrhythmogenic right ventricular cardiomyopathy type 1. *Cardiovasc. Res.* **2005**, *65*, 366–373.
218. Merner, N.D.; Hodgkinson, K.A.; Haywood, A.F.M.; Connors, S.; French, V.M.; Drenckhahn, J.D.; Kupprion, C.; Ramadanova, K.; Thierfelder, L.; McKenna, W.; et al. Arrhythmogenic right ventricular cardiomyopathy type 5 is a fully penetrant, lethal arrhythmic disorder caused by a missense mutation in the *tmem43* gene. *Am. J. Hum. Genet.* **2008**, *82*, 809–821. [[CrossRef](#)] [[PubMed](#)]
219. Taylor, M.R.G.; Slavov, D.; Gajewski, A.; Vlcek, S.; Ku, L.; Fain, P.R.; Carniel, E.; Di Lenarda, A.; Sinagra, G.; Boucek, M.M.; et al. Thymopoietin (lamina-associated polypeptide 2) gene mutation associated with dilated cardiomyopathy. *Hum. Mutat.* **2005**, *26*, 566–574. [[CrossRef](#)] [[PubMed](#)]
220. Kimura, A.; Harada, H.; Park, J.E.; Nishi, H.; Satoh, M.; Takahashi, M.; Hiroi, S.; Sasaoka, T.; Ohbuchi, N.; Nakamura, T.; et al. Mutations in the cardiac troponin i gene associated with hypertrophic cardiomyopathy. *Nat. Genet.* **1997**, *16*, 379–382. [[PubMed](#)]
221. Murphy, R.T.; Mogensen, J.; Shaw, A.; Kubo, T.; Hughes, S.; McKenna, W.J. Novel mutation in cardiac troponin i in recessive idiopathic dilated cardiomyopathy. *Lancet* **2004**, *363*, 371–372. [[CrossRef](#)]
222. Kamisago, M.; Sharma, S.D.; DePalma, S.R.; Solomon, S.; Sharma, P.; McDonough, B.; Smoot, L.; Mullen, M.P.; Woolf, P.K.; Wigle, E.D.; et al. Mutations in sarcomere protein genes as a cause of dilated cardiomyopathy. *N. Engl. J. Med.* **2000**, *343*, 1688–1696. [[CrossRef](#)] [[PubMed](#)]
223. Thierfelder, L.; Watkins, H.; Macrae, C.; Lamas, R.; McKenna, W.; Vosberg, H.P.; Seidman, J.G.; Seidman, C.E. Alpha-tropomyosin and cardiac troponin-t mutations cause familial hypertrophic cardiomyopathy—A disease of the sarcomere. *Cell* **1994**, *77*, 701–712. [[CrossRef](#)]
224. Sibbing, D.; Pfeufer, A.; Perisic, T.; Mannes, A.M.; Fritz-Wolf, K.; Unwin, S.; Sinner, M.F.; Gieger, C.; Gloeckner, C.J.; Wichmann, H.E.; et al. Mutations in the mitochondrial thioredoxin reductase gene *txnr2* cause dilated cardiomyopathy. *Eur. Heart J.* **2011**, *32*, 1121–1133. [[CrossRef](#)] [[PubMed](#)]
225. Gerull, B.; Gramlich, M.; Atherton, J.; McNabb, M.; Trombitas, K.; Sasse-Klaassen, S.; Seidman, J.G.; Seidman, C.; Granzier, H.; Labeit, S.; et al. Mutations of *ttn*, encoding the giant muscle filament titin, cause familial dilated cardiomyopathy. *Nat. Genet.* **2002**, *30*, 201–204. [[CrossRef](#)] [[PubMed](#)]
226. Taylor, M.; Graw, S.; Sinagra, G.; Barnes, C.; Slavov, D.; Brun, F.; Pinamonti, B.; Salcedo, E.E.; Sauer, W.; Pyxaras, S.; et al. Genetic variation in titin in arrhythmogenic right ventricular cardiomyopathy-overlap syndromes. *Circulation* **2011**, *124*, 876–885. [[CrossRef](#)] [[PubMed](#)]
227. Olson, T.M.; Illenberger, S.; Kishimoto, N.Y.; Huttelmaier, S.; Keating, M.T.; Jockusch, B.M. Metavinculin mutations alter actin interaction in dilated cardiomyopathy. *Circulation* **2002**, *105*, 431–437. [[CrossRef](#)] [[PubMed](#)]
228. Vasile, V.C.; Ommen, S.R.; Edwards, W.D.; Ackerman, M.J. A missense mutation in a ubiquitously expressed protein, vinculin, confers susceptibility to hypertrophic cardiomyopathy. *Biochem. Biophys. Res. Commun.* **2006**, *345*, 998–1003. [[CrossRef](#)] [[PubMed](#)]
229. Li, X.P.; Luo, R.; Mo, X.Y.; Jiang, R.J.; Kong, H.; Hua, W.; Wu, X.S. Polymorphism of *zbtb17* gene is associated with idiopathic dilated cardiomyopathy: A case control study in a han chinese population. *Eur. J. Med. Res.* **2013**, *18*. [[CrossRef](#)] [[PubMed](#)]

